# CORRESPONDENCE





# BNIP3 as a potential biomarker for the identification of prognosis and diagnosis in solid tumours

Qin Yu<sup>1†</sup>, Wenhao Fu<sup>2†</sup>, Yutang Fu<sup>2†</sup>, Wenjing Ye<sup>2†</sup>, Huiqiong Yan<sup>3,4†</sup>, Zecheng Yu<sup>2</sup>, Ruirui Li<sup>2</sup>, Yili Cai<sup>5</sup>, Yuxin Chen<sup>2</sup>, Lingyun Wang<sup>6</sup>, Xianqiao Wei<sup>2</sup>, Yangkun Chen<sup>2</sup>, Yuheng Zhang<sup>2</sup>, Huazhong Ying<sup>3,4,7</sup>, Furong Tang<sup>8,9\*</sup>, Fangwei Dai<sup>3,4,7\*</sup> and Wei Han<sup>3,4,7\*</sup>

# Abstract

**Background** Traditional radiotherapy and chemotherapy have been intensively studied for their role in the treatment of tumours. However, these therapies often cause side effects for patients, which calls for the development of novel treatment options for tumours. B-cell lymphoma-2 (Bcl-2)/adenovirus E1B 19 kDa-interacting protein 3 (BNIP3) reportedly apoptosis-inducing effects in tumour cells and is associated with the progression and treatment of multiple tumours. Nevertheless, little is known about its potential role in tumour diagnosis and targeted therapy.

**Findings** The results of the study demonstrated that the interaction of BNIP3 with HDAC1 may affect the progression of breast invasive cancer (BRCA), sarcoma (SARC), kidney renal clear cell carcinoma (KIRC), and low-grade glioma (LGG). BNIP3 seemed to exert its effects in BRCA and SARC primarily through gene silencing and integrator complex, and in KIRC and LGG, mainly by affecting olfactory function, suggesting that targeted therapy can be developed based on the above signalling pathway and downstream molecules.

**Interpretation** BNIP3 has emerged as a promising therapeutic and diagnostic target for BRCA, SARC, KIRC, and LGG, providing new insights into tumour molecular therapies in the clinic.

Keywords BNIP3, Immune infiltration, Pan-cancer analysis, Prognostic biomarker, Diagnosis

<sup>†</sup>Qin Yu, Wenhao Fu, Yutang Fu, Wenjing Ye and Huiqiong Yan contributed equally as the first author.

\*Correspondence: Furong Tang furong.tang@hotmail.com Fangwei Dai fangweidai@163.com Wei Han hanwei3612@163.com <sup>1</sup>School of Information Engineering, Hangzhou Medical College, Hangzhou, Zhejiang, China <sup>2</sup>School of Medical Imaging, Hangzhou Medical College, Hangzhou, Zhejiang, China  <sup>3</sup>Center of Laboratory Animal, Hangzhou Medical College, Hangzhou 310013, Zhejiang, China
<sup>4</sup>Zhejiang Provincial Key Laboratory of Laboratory Animals and Safety Research, Hangzhou Medical College, Hangzhou 310013, Zhejiang, China
<sup>5</sup>School of Clinical Medicine, Hangzhou Medical College, Hangzhou, Zhejiang, China
<sup>6</sup>School of Medical Laboratory and Biological Engineering, Hangzhou Medical College, Hangzhou, Zhejiang, China
<sup>7</sup>Engineering Research Center of Novel Vaccine of Zhejiang Province, Hangzhou Medical College, Hangzhou 310013, Zhejiang, China
<sup>8</sup>The Quzhou Affiliated Hospital of Wenzhou Medical University, Quzhou People's Hospital, Quzhou, Zhejiang, China
<sup>9</sup>Department of Basic Medical Sciences, School of Medicine, Tsinghua University, Beijing, China



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

As a malignant disease, cancer is a serious global public health issue that has permanently affected human society. Cancer remains the leading cause of death in China despite the improvement in health care and increases in funding for cancer control over recent years [1]. Unfortunately, the immune system of cancer patients is generally significantly compromised and altered after long and painful conventional treatments, such as chemotherapy and radiotherapy [2]. As a result, attention is shifting to emerging immune cell therapies because of their advantages of targeting, high efficiency, gene editing, and few side effects. The current study analysed the potential of -cell lymphoma-2 (Bcl-2)/adenovirus E1B 19 kDa-interacting protein 3 (BNIP3) as a potential prognostic marker for tumours.

BNIP3 is a mitochondrial pro-apoptotic protein in the Bcl-2 superfamily, which is vital for the induction of tumour cell apoptosis, as well as the regulation of mitochondrial autophagy, apoptosis, and other functions in tumour cells. Of note, Rossi et al. demonstrated that BNIP3 modulated melanoma development through gene silencing and autophagy [3]. Gorbunova et al. also observed that BNIP3 promoted tumour growth in lung cancer by reducing cell autophagy [4]. Common treatments for cancers, particularly breast invasive cancer (BRCA), sarcoma (SARC), kidney renal clear cell carcinoma (KIRC), and low-grade glioma (LGG), include surgical resection, radiotherapy, chemotherapy, targeted therapy, and immunotherapy. Studies have shown that BNIP3 can affect the therapeutic outcomes of radiotherapy and chemotherapy, as well as serve as a biological marker for targeted therapy, in the treatment research for these four cancers [5]. Nonetheless, the molecular mechanisms of BNIP3 in tumour therapies are poorly understood.

Accordingly, the present study used a multi-omics database and R language to conduct differential, survival, clinical correlation, microsatellite instability (MSI), and correlation analyses of BNIP3 expression in 33 pan-cancerous tumours, explicating the tumour mutational burden (TMB), tumour microenvironment, and immune cell infiltration. The results showed that BNIP3 expression in tumour samples was markedly lower than that in normal samples and that the low expression of BNIP3 was associated with the poor prognosis of BRCA, SARC, KIRC, and LGG. To investigate the molecular mechanism of BNIP3 in these tumours, the signalling pathways of BNIP3 were further analysed through protein-protein interaction (PPI) and gene set enrichment analyses (GSEA). Subsequently, we explored treatments that target BNIP3 by studying the molecules involved in its signalling pathways to provide new ideas for tumour diagnosis and treatment.

## **Results and discussion**

Transcriptomic, mutation, and clinical cancer data were downloaded from the UCSC Xena (http://xena.ucsc. edu/) database containing 10,327 cancer samples and 730 normal samples, which yielded 11,057 transcriptomic data. We used the R language to perform gene ID conversion on the downloaded gene data, extract the BNIP3 gene from the transcriptome data, and form tables of sample names, gene expression, sample types, and tumour types. The data were also analysed in series using R packages limma, ggpubr, etc. There were no available samples for some cancer types. Among these 33 cancers, 23 exhibited low BNIP3 expression, whereas 10 had extremely significantly high expression of BNIP3 (Fig. 1A), indicating that the abnormal expression of BNIP3 may be involved in the development of multiple malignancies. In the prognostic analysis with the forest plot (Fig. 1B, C), low BNIP3 expression was correlated with overall analysis (OS) in 10 tumours and disease-specific survival (DSS) in 7 tumours, illustrating the prognostic significance of BNIP3.

The clinical data of individual cancers were utilised for analysis, and the results of Kaplan-Meier survival curves (Fig. 1D-K) exhibited that poor prognosis was associated with BINP3 upregulation in patients with BRCA (P=0.017) and SARC (P<0.001) and with BNIP3 downregulation in patients with KIRC (P=0.005) or LGG (P<0.001). BNIP3 expression varied by stage, age, and sex (Fig. 1L). As previously reported, more pronounced TMB and MSI suggest that tumour cells produce more neoantigens that can be recognised by immune cells as foreign substances, thus inducing anti-tumour immune responses [6]. In BRCA and KIRC, BNIP3 expression was significantly positively correlated with TMB and MSI, that is, patients with higher BNIP3 expression in this tumour also had higher TMB and longer OS. By contrast, BNIP3 expression was negatively and positively correlated with TMB and MSI, respectively, in LGG.

A prior study reported that TMB was correlated with DNA damage/repair pathways and that higher TMB levels represented higher levels of DNA damage [7]. Therefore, correlation analyses of BNIP3 expression with tumour microenvironment and immune cell infiltration were conducted. In the correlation analysis of the tumour microenvironment, the correlation of BNIP3 expression with immune and stromal cell levels was positive in BRCA (r = -0.22, r = -0.29), SARC (r = -0.17, r =-0.18), LGG (r = -0.39, r = -0.31) but negative in KIRC (r = -0.061, r = -0.1) (Fig. 1O–V), implicating that BNIP3 upregulation was associated with lower immune and stromal cell levels and higher tumour cell purity, which could lead to shorter survival of patients [8]. Regarding immune cell infiltration, BNIP3 expression was substantially correlated with immune cells, especially mast



Fig. 1 BNIP3 expressed prognostic clinical tumour biology in pan-cancer, and immune microenvironments in BRCA, SARC, KIRC, and LGG. A: BNIP3 mRNA expression in different types of cancer. B-C: COX regression analysis. D-K: Kaplan-Meier survival analysis. L: The relationship between BNIP3 expression and three clinical characteristics (age, gender, and stage) in 33 types of cancers. M-N: Correlation between BNIP3 and MSI. O-Z: The relationship between BNIP3 expression and different types of immune cell infiltration levels in cancers



Fig. 2 GSEA and Venn diagram and PPI gene co-expression mining BNIP3 as a diagnostic molecular marker for BRCA, SARC, KIRC, and LGG. A: Signalling pathways associated with BNIP3 expression in SARC. C: Signalling pathways associated with BNIP3 expression in SARC. C: Signalling pathways associated with BNIP3 expression in KIRC. D: Signalling pathways associated with BNIP3 expression in LGG. E: Venn diagram of KIRC and LGG. F: Venn diagram of KIRC and LGG. F: Venn diagram of KIRC and LGG. G: The intersect pathways between SARC and BRCA and between KIRC and LGG. H-I: Correlation between BNIP3 expression and related gene (immune checkpoint genes and interacted genes) expression. J: PPI network for BNIP3-interacting genes

cells, in BRCA (r = -0.142, P<0.001), SARC (r = -0.27, P<0.001), and LGG (r = -0.166, P=0.004). Mast cells, innate immune cells in tissues, are important for tissue homeostasis and inflammation. Mast cell proliferation shares an association with certain pathological conditions, and mast cells are associated with several health and disease states [9]. Our analysis also confirmed that BNIP3 expression modulated mast cells and the tumour microenvironment, thus affecting patient survival.

BNIP3 exerts its effects on the occurrence of different types of cancer through various biological processes. We explored potential pathways and genes through which BNIP3 may affect tumourigenesis of the four tumours using the GSEA and Venn diagram analysis. The results showed that BNIP3 regulated the tumourigenesis of BRCA and SARC primarily through six pathways, including ncRNA 3' end processing, snRNA processing, integrator complex, snRNA 3' end processing, snRNA metabolic process, and gene silencing (Fig. 2A-G). Further, BNIP3 affects the tumorigenesis of KIRC and LGG mainly via UTR mediated mRNA, positive regulation of translational initiation, regulation of translational initiation, U2 snRNP, regulation of artery morphogenesis, presynaptic active zone cytoplasmic component, inhibitory extracellular ligand-gated ion channel activity, translation activator activity, benzodiazepine receptor activity, and ligand-gated anion channel activity pathways.

PPI analyses were further conducted to probe the molecular mechanism of action of BNIP3 in tumours. The PPI network revealed that an interaction existed between BNIP3 and histone deacetylases (HDACs), which are involved in several core processes regulating endothelial cell physiological functions, especially angiogenesis, inflammatory signalling, and redox homeostasis (Fig. 2H-I). Most endothelial HDACs have anti-proliferative functions, among which class I HDACs promote the proliferation of cancer cells. In addition to different external stimuli, external stressors also affect whether HDAC1 functions in pro- or anti-inflammatory pathways [10, 11]. This fact also explains, to some extent, the phenomenon that the high expression of BNIP3 in different tumours plays different roles in tumour metastasis [12], which is one of the major causes of death in cancer patients. Therefore, we hypothesised that BNIP3 also interacts with HDACs to contribute to cancer cell proliferation and metastasis.

In KIRC and LGG, the prognosis of patients with low BNIP3 expression was poorer than that of patients with high expression (Fig. 1B–C), and BNIP3 expression varied between men and women in different subgroups of KIRC (Fig. 1L). HDACs are responsible for neuronal survival [13], and BNIP3 can maintain intra-mitochondrial homeostasis by degrading damaged mitochondria through mitochondrial phagocytosis and is implicated in renal cell survival and renal function stability [14, 15]. How BNIP3 affects the olfactory pathway, leading to changes in olfactory function, will be further explored in future studies as a diagnostic basis for the aforementioned four types of cancer.

Alterations in olfactory function can be affected by various factors, including environmental factors and variations. Therefore, biological correlation studies did not imply any causal relationship. Meanwhile, as a necessary condition for developing a detection method in tumour and eliminating inequality in cancer treatment, this study needs to encompass a broader population and include a wider range of cancer types, including less prevalent ones. Future validation of BNIP3 in large sample sizes is warranted prior to its clinical application as a diagnostic biomarker.

# In conclusion

BNIP3 exhibited abnormal expression in 33 tumours, which is associated with poor prognosis in BRCA, SARC, KIRC, and LGG. Clinical tumour prognosis evaluation through the expression of BNIP3 in BRCA, SARC, KIRC, and LGG revealed a potential for favourable immunotherapeutic efficacy of BNIP3 in these tumours. Furthermore, a correlation analysis was conducted on immune microenvironment indicators. Multi-omics GSEA and other approaches indicateds that BNIP3 interacted with HDAC1 and affected tumour prognosis by influencing olfactory function, suggesting that targeted regulation of olfactory function by BNIP3 and HDAC1 may serve as a potential clinical therapy prognostic diagnosis in BRCA, SARC, KIRC, and LGG.

#### Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12943-023-01808-9.

Supplementary Material 1: **Fig. 1**: Among the four types of tumours (BRCA, SARC, KIRC, and LGG), the correlation between the three clinical features and BNIP3 expression was significant.

Supplementary Material 2: **Table 1**: Total data of differential expression analysis and survival analysis of BNIP3 in 33 types of tumours.

Supplementary Material 3: **Table 2:** The full names and abbreviations of 33 types of tumours correspond to the table.

#### Acknowledgements

The authors gratefully thank (Key Discipline of Zhejiang Province in Public and Preventive Medicine (First Class, Category A), Hangzhou Medical College) for their support.

#### Authors' contributions

QY wrote the text of the paper and produced all the figures, WHF, YTF, WJY, QHY provided all the data and results of the research analysis map.ZCY, RRL, YLC, YXC, LYW, XQW, YKC, YHZ, HZY contributed to study design, multi-omics analysis method support, biostatistical difference analysis, and manuscript revision.FRT provided technical guidance. FRT, FWD, and WH provided comments on the writing of the thesis and made changes. WH collected

#### Funding

This Work is supported by National Natural Science Foundation of China (grant no: 62271353, 62001311), Natural Science Foundation of Sichuan Province (grant no: 2022NSFSC0926), Fellowship of China Postdoctoral Science Foundation (grant no: 2023M731984), Hangzhou Medical College Institute Special Project (grant no: YS2021009), Department of Education of Zhejiang Province (grant no: Y201942573), Medical Science and Technology Project of Zhejiang Province (grant no: 2018KY351, 2020KY530), Zhejiang Traditional Chinese Medicine Administration (grant no: 2021ZB081), The Natural Science Foundation of Zhejiang Province (grant no: LXZ22H300001) and National Innovation and Entrepreneurship Training Program for College Students (grant no: 202013023018, 20231302303).

## Data Availability

The data supporting this study's findings are openly available in Xena Ucsc at http://xena.ucsc.edu/. The data supporting this study's findings are openly available in Bioconductor at http://master.bioconductor.org/. The data supporting this study's findings are openly available in String at http:// cn.string-db.org/. The data supporting this study's findings are openly available in Evenn at http://www.ehbio.com/test/venn/#/.

### Declarations

Ethics approval and consent to participate Not applicable.

#### **Consent for publication**

Not applicable.

#### Competing interests

The authors declare no competing interests.

Received: 23 May 2023 / Accepted: 17 June 2023 Published online: 30 August 2023

#### References

1. Cao M, Li H, Sun D, Chen W. Cancer burden of major cancers in China: a need for sustainable actions. Cancer Commun (Lond). 2020;40:205–10.

- Han HJ, Nwagwu C, Anyim O, Ekweremadu C, Kim S. COVID-19 and cancer: from basic mechanisms to vaccine development using nanotechnology. Int Immunopharmacol. 2021;90:107247.
- Vara-Pérez M, Rossi M, Van den Haute C, Maes H, Sassano ML, Venkataramani V, et al. BNIP3 promotes HIF-1α-driven melanoma growth by curbing intracellular iron homeostasis. Embo J. 2021;40:e106214.
- Gorbunova AS, Yapryntseva MA, Denisenko TV, Zhivotovsky B. BNIP3 in Lung Cancer: to kill or rescue? Cancers (Basel). 2020;12:3390.
- Choi GE, Lee HJ, Chae CW, Cho JH, Jung YH, Kim JS, Kim SY, Lim JR, Han HJ. BNIP3L/NIX-mediated mitophagy protects against glucocorticoid-induced synapse defects. Nat Commun 2021 Jan 20;12(1):487.
- Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science. 2015;348:124–8.
- 7. ZHANG J, LIU M, FANG Y, et al. TP53 R273C mutation is Associated with Poor Prognosis in LGG patients [J]. Front Genet. 2022;13:720651.
- Zhang J, Liu M, Fang Y, Li J, Chen Y, Jiao S. TP53 R273C mutation is Associated with Poor Prognosis in LGG Patients. Front Genet. 2022;13:720651.
- da Silva EZ, Jamur MC, Oliver C. Mast cell function: a new vision of an old cell. J Histochem Cytochem. 2014;62:698–738.
- Dunaway LS, Pollock JS. HDAC1: an environmental sensor regulating endothelial function. Cardiovasc Res. 2022;118:1885–903.
- 11. Yoon S, Eom GH. HDAC and HDAC inhibitor: from Cancer to Cardiovascular Diseases. Chonnam Med J. 2016;52:1–11.
- 12. Lin Y, Xu J, Lan H. Tumor-associated macrophages in tumor metastasis: biological roles and clinical therapeutic applications. J Hematol Oncol. 2019;12:76.
- Chen JS, Wang HK, Hsu CY, Su YT, Chen JS, Liang CL, et al. HDAC1 deregulation promotes neuronal loss and deficit of motor function in stroke pathogenesis. Sci Rep. 2021;11:16354.
- Bhatia D, Chung KP, Nakahira K, Patino E, Rice MC, Torres LK, et al. Mitophagydependent macrophage reprogramming protects against kidney fibrosis. JCI Insight. 2019;4:e132826.
- Lin Q, Li S, Jiang N, Jin H, Shao X, Zhu X, et al. Inhibiting NLRP3 inflammasome attenuates apoptosis in contrast-induced acute kidney injury through the upregulation of HIF1A and BNIP3-mediated mitophagy. Autophagy. 2021;17:2975–90.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.