

Review

Open Access

Pathways for aberrant angiogenesis in pancreatic cancer

M Korc*

Address: Division of Endocrinology, Diabetes, and Metabolism, Departments of Medicine, Biological Chemistry, and Pharmacology, University of California, Irvine, California 92697, USA

Email: M Korc* - mkorc@uci.edu

* Corresponding author

Published: 7 January 2003

Received: 9 December 2002

Molecular Cancer 2003, 2:8

Accepted: 7 January 2003

This article is available from: <http://www.molecular-cancer.com/content/2/1/8>

© 2003 Korc; licensee BioMed Central Ltd. This is an Open Access article: verbatim copying and redistribution of this article are permitted in all media for any purpose, provided this notice is preserved along with the article's original URL.

Abstract

Pancreatic ductal adenocarcinoma (PDAC) is a devastating disease. Although the specific mechanisms that dictate its biological aggressiveness are not clearly established, it is characterized by a variety of molecular alterations as well as by the overexpression of mitogenic and angiogenic growth factors and their receptors. PDACs also express high levels of vascular endothelial growth factor (VEGF). Recent studies indicate that suppression of VEGF expression attenuates pancreatic cancer cell tumorigenicity in a nude mouse model, and that VEGF can exert direct mitogenic effects on some pancreatic cancer cells. These findings suggest that cancer cell derived VEGF promotes pancreatic cancer growth in vivo via a paracrine angiogenic pathway and an autocrine mitogenic pathway, and provide novel opportunities for therapeutic intervention in this deadly disease.

Carcinoma of the pancreas: An overview

Pancreatic ductal adenocarcinoma (PDAC) is responsible for over 20% of deaths due to gastrointestinal malignancies, making it the fourth most common cause of cancer related mortality in the United States and other industrialized countries. The prognosis of patients with PDAC is extremely poor, with overall 5-year survival rates that are less than 1% [1], one-year overall survival of 12%, and a median survival of 6 months [2]. Survival is often limited to patients who had surgical resection at an early stage of the disease. However, the diagnosis of PDAC is often established at an advanced stage, precluding patients from undergoing tumor resection in spite of limited results with other treatment modalities [3]. These dismal statistics are due to the tumor's propensity to metastasize when small and undetectable, the advanced stage at which many patients first develop symptoms, and the intrinsic resistance of pancreatic cancer cells to cytotoxic agents and radiotherapy [3–5]. PDAC may be an even more serious problem in the future since its incidence increases after age 50 and the general population world-wide is aging.

There is, therefore, an urgent need for an improved understanding of the mechanisms that contribute to pancreatic tumor growth and metastasis, and for the design of therapies for this disorder that are more effective than current regimens. This review will cover in a brief manner the molecular biology of pancreatic cancer, and will then focus on various aspects of vascular endothelial growth factors in angiogenesis in general and in relation to PDAC in particular.

Molecular biology of pancreatic cancer

A plethora of genetic mutations have been described in the cancer cells of PDAC patients. The most frequent alterations (approximate frequency indicated in parenthesis) include mutations in the *K-ras* oncogene (90%), the p53 (85%) and Smad4 (50%) tumor suppressor genes, and the p16 (85% mutated and 15% silenced epigenetically) cell cycle inhibitory gene [6,7]. Together, these alterations promote cellular proliferation, suppress apoptotic pathways, and facilitate tumor spread and metastasis. In addition, there is overexpression of multiple tyrosine kinase

receptors and their ligands which enhances mitogenesis, and loss of responsiveness to the growth-inhibitory signals of members of the transforming growth factor beta (TGF- β) family [6,7], which contribute in a significant manner to the biological aggressiveness of PDAC.

It is well established that human pancreatic cancer cell lines overexpress the epidermal growth factor (EGF) receptor (EGFR) and produce multiple ligands that bind directly to EGFR, including transforming growth factor- α (TGF- α), amphiregulin, heparin-binding EGF-like growth factor (HB-EGF), betacellulin and epiregulin [8–12]. These cell lines also express other growth factors such as fibroblast growth factors (FGFs) and platelet-derived growth factor (PDGF) B chain [13–16]. However, expression of receptors and ligands in cell lines does not necessarily indicate parallel alterations in PDAC *in vivo*. Therefore, studies using human tissues have been of vital importance in this regard. Studies using immunohistochemistry, Northern blot analysis and *in situ* hybridization techniques, have demonstrated that PDAC tissue samples overexpress EGFR and six ligands that bind directly to EGFR (EGF, TGF- α , HB-EGF, betacellulin, epiregulin and amphiregulin), as well as c-erb-B2, c-erb-B3, and c-erb-B4 [10,11,17–19]. These cancers also overexpress basic fibroblast growth factor (FGF-2), acidic FGF (FGF-1), keratinocyte growth factor (KGF), FGF-5, PDGF B chain (but not A chain), insulin-like growth factor-I (IGF-I), the EGF-like growth factor Cripto, hepatocyte growth factor (HGF), vascular endothelial growth factor (VEGF), all 3 mammalian transforming growth factor beta (TGF- β) isoforms, bone morphogenetic protein-2 (BMP-2) and activin β A [14,15,20–29]. Many, but not all of the corresponding receptors are concomitantly overexpressed. For example, there is overexpression of PDGF receptor α and β , the IGF-1 receptor, MET (the receptor that binds HGF), the 2 Ig-like form of type I FGF receptor (FGFR-1), and the type II TGF- β receptor (T β RII) but not the insulin receptor [16,21,23,26,30–33]. IGF-II and insulin are not overexpressed in PDAC [21], whereas the type I TGF- β receptor (T β RI) is under-expressed [31–33]. Thus, there is selective overexpression of specific receptors and their ligands in PDAC, and this concomitant overexpression leads to the creation of aberrant paracrine and autocrine pathways that confers a distinct growth advantage to pancreatic cancer cells.

The clinical importance of the above observations is underscored by numerous observations. For example, the concomitant presence in the cancer cells of EGFR and either EGF or TGF- α is associated with disease progression and decreased survival of PDAC patients [34]. Overexpression of c-erbB3 [19], FGF-2 [20] or TGF- β [35] is associated with decreased patient survival. The aberrant cytoplasmic localization of amphiregulin [36] is also as-

sociated with decreased patient survival. Dominant negative inhibition of either EGFR or FGFR-1 markedly attenuates pancreatic cancer cell growth [37–39]. Expression of a cyclin D1 antisense construct in pancreatic cancer cells lowers cyclin D1 levels in these cells, attenuates their growth *in vitro*, and blocks their tumorigenicity *in vivo* [40]. EGFR blockade with an anti-EGFR antibody attenuates pancreatic tumor growth, and inhibition of EGFR tyrosine kinase activity suppresses pancreatic tumor angiogenesis [41,42]. Together, these findings are among many that support the hypothesis that tyrosine kinase receptors and ligands have an important role in PDAC.

VEGF family of growth factors and their receptors

VEGF-A, also called "vascular permeability factor", is a homodimeric heparin-binding glycoprotein [43–45]. Five major VEGF-A isoforms having 121, 145, 165, 189 and 206 amino acid residues, respectively, arise as a result of alternative splicing from a single gene [46,47]. VEGF-A₁₂₁ and VEGF-A₁₄₅ are usually secreted while VEGF-A₁₈₉ and VEGF-A₂₀₆ are almost completely sequestered in the extracellular matrix [47]. VEGF-A₁₆₅ is half secreted and half bound to the cell surface and the extracellular matrix [48]. All 5 isoforms are mitogenic toward vascular endothelial cells and induce vascular permeabilization. Additional VEGF isoforms and VEGF-related genes have been identified, including VEGF-B [49,50], VEGF-C [51], VEGF-D [52], VEGF-E [53] and placenta growth factor [54]. Direct evidence for the role played by VEGF-A in embryonic vasculogenesis and angiogenesis was also demonstrated in VEGF-A gene knockout studies [55,56], in which loss of a single VEGF-A allele in mice resulted in embryonic lethality between day 11 and 12. Angiogenesis and blood-island formation were impaired, resulting in severe developmental anomalies. This heterozygous lethal phenotype is indicative of the tight dose-dependent regulation of embryonic vessel development by VEGF-A [55,56]. VEGF-A is also required for the cyclical blood vessel proliferation in the female reproductive tract and for longitudinal bone growth and endochondral bone formation in postnatal development [43]. Together, these observations indicate that VEGF-A has an important role in embryogenesis, development, and tissue remodeling.

VEGF-A stimulates endothelial cell proliferation through binding to two related tyrosine kinase receptors, VEGFR-1 (flt-1) VEGFR-2 (flk-1/KDR), on the surface of endothelial cells, with most of the mitogenic effects taken to occur via VEGFR-2 [57–59] (57–59). A third high affinity VEGF receptor, termed VEGFR-3 (Flt4), is expressed in lymphatic vessels [60,61]. It is activated by VEGF-C, which can be processed to a form that also binds to VEGFR-2 [57–61]. Furthermore, placenta growth factor and VEGF-B bind only VEGFR-1, whereas VEGF-D, like VEGF-C, interacts

with both VEGFR-2 and VEGFR-3 [57–61]. However, VEGF-E binds only to VEGFR-2 [59]. All three VEGFRs are class III transmembrane protein tyrosine kinases that possess seven immunoglobulin-like sequences in their extracellular domains and a kinase insert in their intracellular domains [57–61]. In addition, neuropilin-1 (Np-1), a neuronal guidance molecule for axons in the developing nervous system, also acts as a co-receptor for VEGF-A₁₆₅ (but not for VEGF-A₁₂₁), PlGF-2, VEGF-B and VEGF-E [62]. Np-1 is a non-tyrosine kinase transmembrane protein whose overexpression in transgenic mice is associated with various abnormalities, including excess capillary and blood vessel formation [63]. The closely related neuropilin-2 (Np-2) also binds VEGF-A₁₆₅ (but not VEGF-A₁₂₁), as well as VEGF-A₁₄₅ and PlGF-2, strongly implying that both Np-1 and Np-2 in angiogenesis [62–64].

Gene knockout studies have shown that both VEGFR-1^{-/-} and VEGFR-2^{-/-} mice die *in utero* between day 8.5 and 9.5 [65,66]. In VEGFR-1^{-/-} mice, endothelial cells developed in both embryonic and extra-embryonic sites but failed to organize into normal vascular channels [65]. In VEGFR-2^{-/-} mice, hematopoietic precursors were severely reduced, yolk-sac blood islands were absent, organized blood vessels failed to develop throughout the embryo or the yolk sac [66]. Furthermore, double knockouts for Np-1 and Np-2 die *in utero* between day 8.5 and 9.5 [67]. They exhibit avascular yolk sacs, and mice that are deficient for Np-1 but heterozygous for Np-2, or deficient for Np-2 but heterozygous for Np-1, die at day 10 to 10.5 and exhibit diffuse vascular abnormalities that are more marked than either Np-1 or Np-2 single knockouts [67]. Together, these observations suggest that VEGFR-1 and VEGFR-2 are essential for embryonic vasculature development, whereas VEGFR-3 is essential for lymphangiogenesis, and that Np-1 and Np-2 are as important as the other components of the VEGF pathway in embryonic angiogenesis.

Angiogenesis in cancer

Tumor angiogenesis is often the consequence of an angiogenic imbalance in which pro-angiogenic factors predominate over anti-angiogenic factors [68–71]. Furthermore, angiogenesis is essential for growth and metastasis of most solid malignancies, and VEGF-A is believed to be critical for tumor angiogenesis [72,73]. Thus, secretion of bioactive VEGF-A by cancer cells may be directly involved in tumor progression [43]. For example, ovarian cancer cells secrete large amounts of bioactive VEGF-A that may play a crucial role in the genesis of ascitic fluid accumulation, angiogenesis and tumor induced immunosuppression in ovarian cancer patients [74]. In high grade gliomas, bioactive VEGF-A secreted by the glioma cells may account for the histopathological and clinical features of these tumors, including such characteristics as

marked tumor angiogenesis and increased cerebral edema [75,76].

VEGF-A expression is induced by multiple mechanisms. These include mutant *K-ras* and mutant *p53*, the von Hippel Lindau gene product, growth factors such as FGF-2 and TGF- β , hypoxia, and transcription factors such as hypoxia inducible factor 1 alpha and SP1 [77–81]. VEGF-A is up-regulated in many tumors including mammary, colorectal, renal, liver, ovarian and gastric carcinomas and gliomas [43], and its overexpression has been correlated with poor prognosis. For example, breast cancer patients with metastatic disease whose tumors exhibit increased angiogenesis have a worse prognosis than the corresponding patients whose tumors do not exhibit increased angiogenesis [82]. Furthermore, suppression of VEGF-A functions inhibits tumor growth in animal models as demonstrated with a dominant negative VEGFR-2, soluble VEGFR-1, neutralizing anti-VEGF-A antibody, VEGF-A anti-sense expression, anti-VEGFR-1 or anti-VEGFR-2 ribozymes, tyrosine kinase inhibitors of VEGFR-2, and anti-VEGFR-2 antibodies [83–92].

Role of VEGF in pancreatic cancer angiogenesis

Although PDAC is not a grossly vascular tumor, this malignancy often exhibits enhanced foci of endothelial cell proliferation. Moreover, several [24,93,94], but not all [95] studies, have reported a positive correlation between blood vessel density, tumor VEGF-A levels, and disease progression in PDAC, raising the possibility that VEGF-A may have an important role in this disease. However, PDACs overexpress multiple additional mitogenic growth factors which are also angiogenic (Table 1), such as EGF, TGF- α , HGF, FGFs such as FGF-1, FGF-2, and FGF-5, and PDGF-beta [6,96]. Therefore, while VEGF-A is of crucial importance in promoting the growth and metastasis of pancreatic cancer cells in PDAC, other factors are most likely also involved in this process. Nonetheless, it has been demonstrated that pancreatic cancer cells secrete biologically active VEGF-A [25], and the cancer cells in PDAC as well as pancreatic cancer cell lines sometimes express VEGFR-1 and/or VEGFR-2 [97]. Moreover, some of these cells may be growth stimulated by VEGF-A in cell culture [97,98], and the major angiogenic agent toward human dermal microvascular endothelial cells (HDMEC) that is produced by T3M4 and PANC-1 human pancreatic cancer cells is VEGF-A, since the mitogenic activity of conditioned medium from these cells can be nearly completely suppressed by neutralizing anti-VEGF-A antibodies [99]. Together, these observations suggest that by promoting angiogenesis VEGF-A enhances tumor spread and metastasis in this malignancy.

In support of the above conclusion, it has been demonstrated that anti-angiogenic therapy is effective at sup-

Table 1: Examples of Angiogenic Growth Factors that Are Overexpressed in Human Pancreatic Cancer and their Cognate Receptors

| Growth Factors Activating Tyrosine Kinase Receptors | Receptor |
|--|-------------------------------------|
| VEGF-A | VEGFR-1 and VEGFR-2 |
| VEGF-C | VEGFR-3 |
| EGF, TGF- α , HB-EGF | EGF receptor |
| FGF-1, -2, -5 | FGF receptors, types 1 and 2 |
| PDGF B chain | PDGF receptors α and β |
| IGF-I | IGF-I receptor |
| Hepatocyte growth factor | MET |
| Growth Factors that Activate Serine-Threonine Kinase Receptors | |
| TGF- β 1, -2, -3 | Type II TGF- β receptor |
| Pro-Angiogenic Chemokines | |
| IL-8 | CXCR1 and CXCR2 |
| Mip 3 α | CCR6 |

pressing tumor growth in animal models of PDAC. Thus, the anti-angiogenic agent TNP-470 reduces neoangiogenesis in tumors formed by pancreatic cancer cell lines, and decreases tumor growth and metastasis [99]. Suppression of VEGF-A expression with a VEGF-A antisense construct and with a VEGF directed ribozyme markedly attenuates tumorigenicity in nude mice and formation of hepatic metastases [25,100]. VEGF-A fused to diphtheria toxin (DT-VEGF) internalizes in target cells via VEGFRs, inhibits protein synthesis, and suppresses the growth of HUVEC endothelial cells, thereby decreasing the volume and microvessel density in tumors formed by pancreatic cancer cells [101]. Adenoviral vectors carrying sequences encoding soluble VEGFR-1 and VEGFR-2 [102,103], or the VEGFR tyrosine kinase inhibitor PTK 787 [104], also inhibit the growth of growth and/or metastasis of pancreatic cancers in mouse models. These findings underscore the importance of the angiogenic process in PDAC, support the hypothesis that VEGF-A exerts a crucial role in this regard, and raise the possibility that VEGF-A may exert direct effects on pancreatic cancer cells *in vivo*.

VEGF-A can also act as a survival factor for endothelial cells, rendering these cells more radioresistant [105]. It can also promote the survival of leukemic cells, certain tumor cells and hematopoietic stem cells [106–108]. In addition, VEGF-C is also overexpressed in PDAC, and this overexpression has been correlated with enhanced lymph node metastasis [109]. Thus, various members of the VEGF family of ligands may contribute to the growth and metastasis of pancreatic cancer cells through a variety of mechanisms.

Additional mechanisms for promoting pancreatic cancer angiogenesis

Although VEGF appears to be of paramount importance for the angiogenic process in PDAC, these cancers express

many other pro-angiogenic factors (Table 1). As in the case of VEGF, some of these growth factors activate tyrosine kinase receptors that are expressed in endothelial cells within the pancreatic tumor mass, such as EGFR [17]. The importance of tyrosine kinase receptors other than VEGFR in pancreatic cancer angiogenesis is underscored by recent observations that inhibition of EGFR tyrosine kinase activity suppresses pancreatic tumor angiogenesis [42], and that NK4, an antagonist that is composed of the N-terminal hairpin and subsequent four-kringle domains of HGF, is a competitive antagonist for HGF that potently inhibits angiogenesis in tumors formed by SUIT-2 pancreatic cancer cells [110].

Other pro-angiogenic factors that are overexpressed in PDAC include certain chemokines such as Mip3 α and interleukin-8 (IL-8), which activate G-protein coupled receptors [111–113]. By contrast, TGF- β s activate serine-threonine kinase receptors [114]. The importance of TGF- β s are pro-angiogenic factors in PDAC is underscored by the recent observation that expression of a soluble T β RII in pancreatic cancer cells interferes with TGF- β actions, attenuates tumor growth and metastasis, and suppresses tumor angiogenesis [Rowland-Goldsmith, 2001 #905; Rowland-Goldsmith MA, 2002 #2548].

Often, there is evidence for cross-talk between the various angiogenic factors. For example, TGF- β 1 and plasminogen activator inhibitor-1 (PAI-1) are overexpressed in PDAC [117,118], TGF- β 1 induces PAI-1 expression in pancreatic cancer cells [119], and both TGF- β 1 and PAI-1 can promote angiogenesis *in vivo* [120–122]. TGF- β s are initially released as latent molecules that form complexes with latent binding protein (LTBP), and their biological effectiveness is dependent on their activation by such proteins as plasmin, uPA and its receptor, the insulin-like growth factor II (IGF-2) receptor, and tissue transglutami-

nase [123,124]. The IGF-2 receptor, as well as uPA and its receptor are overexpressed in PDAC [125,126], and pancreatic cancer cell lines express tissue transglutaminase [127]. Furthermore, uPA and its receptor, as well as tissue transglutaminase, have been implicated in the angiogenic process [128,129], and the angiogenic potential of TGF- β s may be enhanced by the presence of Smad4 mutations [130], which are frequent in PDAC. uPA can transactivate EGFR [131], and EGFR activation can induce the expression of VEGF and the pro-angiogenic chemokine interleukin-8 [132,133]. Taken together, these observations suggest that multiple pathways interact to enhance angiogenesis in PDAC.

The pancreatic microenvironment may also serve to promote tumor angiogenesis [134]. In addition, as a consequence of the existence of a continuous intra-pancreatic portal circulation, pancreatic cancer cells may be exposed to high levels of islet cell derived hormones such as insulin and growth factors such as TGF- β s [135]. High insulin levels bind and activate the IGF-1 receptor, which can then promote angiogenesis [136,137]. Furthermore, islet cell derived TGF- β s may enhance matrix metalloproteinase-9 (MMP-9) and VEGF expression in PDAC [31,138], and suppress PTEN expression [139]. MMP-9 enhances tumor angiogenesis [140] whereas PTEN, a phosphatase with specificity for 3-phosphorylated inositol phospholipids, has been implicated in the suppression of tumor angiogenesis [141].

Conclusion

PDAC is a biologically aggressive malignancy that has a propensity to spread locally and metastasize distally. While not grossly vascular, these cancers exhibit foci of micro-angiogenesis and overexpress multiple pro-angiogenic factors. VEGF and related ligands represent a crucial component of this pro-angiogenic switch, as evidenced by the presence of high levels of VEGF in ascitic fluid of PDAC patients [142], the correlation between high serum VEGF levels and disease recurrence post-operatively [143], and the observation that high VEGFR-2 levels are associated with a worse prognosis in this disease [144]. Therefore, mechanisms that target VEGF and the various pathways that enhance the angiogenic process in PDAC [145] may ultimately be of great therapeutic benefit in patients with unresectable disease as well as following surgery to prevent disease recurrence.

References

- Warshaw AL and Fernandez-del Castillo C **Pancreatic carcinoma.** *N Engl J Med* 1992, **326**(7):455-65
- Parker SL **Cancer statistics, 1997.** *CA Cancer J Clin* 1997, **47**(1):5-27
- Bramhall SR and Neoptolemos JP **Adjuvant chemotherapy in pancreatic cancer.** *Int J Pancreatol* 1997, **21**(1):59-63
- Abrams RA **Role of radiation therapy in the management of the patient with pancreatic cancer.** *Oncology (Huntingt)* 1996, **10**(9 Suppl):13-7
- Kuvshinov BW and Bryer MP **Treatment of resectable and locally advanced pancreatic cancer.** *Cancer Control* 2000, **7**(5):428-36
- Korc M **Role of growth factors in pancreatic cancer.** *Surg Oncol Clin N Am* 1998, **7**(1):25-41
- Kern SE **Molecular genetic alterations in ductal pancreatic adenocarcinomas.** *Med Clin North Am* 2000, **84**(3):691-5
- Korc M, Meltzer P and Trent J **Enhanced expression of epidermal growth factor receptor correlates with alterations of chromosome 7 in human pancreatic cancer.** *Proc Natl Acad Sci U S A* 1986, **83**(14):5141-4
- Smith JJ, Derynck R and Korc M **Production of transforming growth factor alpha in human pancreatic cancer cells: evidence for a superagonist autocrine cycle.** *Proc Natl Acad Sci U S A* 1987, **84**(21):7567-70
- Ebert M **Induction and expression of amphiregulin in human pancreatic cancer.** *Cancer Res* 1994, **54**(15):3959-62
- Kobrin MS **Induction and expression of heparin-binding EGF-like growth factor in human pancreatic cancer.** *Biochem Biophys Res Commun* 1994, **202**(3):1705-9
- Yokoyama YFH, Kobrin MS, Ebert M, Friess H, Büchler MW and Korc M **Betacellulin, a member of the EGF family is overexpressed in human pancreatic cancer.** *Int J Oncol* 1995, **7**:825-829
- Kornmann MH, Beger G and Korc M **Role of fibroblast growth factors and their receptors in pancreatic cancer and chronic pancreatitis.** *Pancreas* 1998, **17**(2):169-75
- Kornmann M **Fibroblast growth factor-5 stimulates mitogenic signaling and is overexpressed in human pancreatic cancer: evidence for autocrine and paracrine actions.** *Oncogene* 1997, **15**(12):1417-24
- Siddiqi I **Increased expression of keratinocyte growth factor in human pancreatic cancer.** *Biochem Biophys Res Commun* 1995, **215**(1):309-15
- Ebert M **Induction of platelet-derived growth factor A and B chains and over-expression of their receptors in human pancreatic cancer.** *Int J Cancer* 1995, **62**(5):529-35
- Korc M **Overexpression of the epidermal growth factor receptor in human pancreatic cancer is associated with concomitant increases in the levels of epidermal growth factor and transforming growth factor alpha.** *J Clin Invest* 1992, **90**(4):1352-60
- Yamanaka Y **Overexpression of HER2/neu oncogene in human pancreatic carcinoma.** *Hum Pathol* 1993, **24**(10):1127-34
- Friess H **Enhanced erbB-3 expression in human pancreatic cancer correlates with tumor progression.** *Clin Cancer Res* 1995, **1**(11):1413-20
- Yamanaka Y **Overexpression of acidic and basic fibroblast growth factors in human pancreatic cancer correlates with advanced tumor stage.** *Cancer Res* 1993, **53**(21):5289-96
- Bergmann U **Insulin-like growth factor I overexpression in human pancreatic cancer: evidence for autocrine and paracrine roles.** *Cancer Res* 1995, **55**(10):2007-11
- Friess H **Cripto, a member of the epidermal growth factor family, is over-expressed in human pancreatic cancer and chronic pancreatitis.** *Int J Cancer* 1994, **56**(5):668-74
- Ebert M **Coexpression of the c-met proto-oncogene and hepatocyte growth factor in human pancreatic cancer.** *Cancer Res* 1994, **54**(22):5775-8
- Itakura J **Enhanced expression of vascular endothelial growth factor in human pancreatic cancer correlates with local disease progression.** *Clin Cancer Res* 1997, **3**(8):1309-16
- Luo J **Pancreatic cancer cell-derived vascular endothelial growth factor is biologically active in vitro and enhances tumorigenicity in vivo.** *Int J Cancer* 2001, **92**(3):361-9
- Friess H **Enhanced expression of the type II transforming growth factor beta receptor in human pancreatic cancer cells without alteration of type III receptor expression.** *Cancer Res* 1993, **53**(12):2704-7
- Kleeff J **Bone morphogenetic protein 2 exerts diverse effects on cell growth in vitro and is expressed in human pancreatic cancer in vivo.** *Gastroenterology* 1999, **116**(5):1202-16
- Kleeff J **Concomitant over-expression of activin/inhibin beta subunits and their receptors in human pancreatic cancer.** *Int J Cancer* 1998, **77**(6):860-8
- Bergmann U **Increased expression of insulin receptor substrate-1 in human pancreatic cancer.** *Biochem Biophys Res Commun* 1996, **220**(3):886-90

30. Kobrin MS **Aberrant expression of type I fibroblast growth factor receptor in human pancreatic adenocarcinomas.** *Cancer Res* 1993, **53(20)**:4741-4
31. Wagner M **Enhanced expression of the type II transforming growth factor-beta receptor is associated with decreased survival in human pancreatic cancer.** *Pancreas* 1999, **19(4)**:370-6
32. Lu Z **Presence of two signaling TGF-beta receptors in human pancreatic cancer correlates with advanced tumor stage.** *Dig Dis Sci* 1997, **42(10)**:2054-63
33. Wagner M **Transfection of the type I TGF-beta receptor restores TGF-beta responsiveness in pancreatic cancer.** *Int J Cancer* 1998, **78(2)**:255-60
34. Yamanaka Y **Coexpression of epidermal growth factor receptor and ligands in human pancreatic cancer is associated with enhanced tumor aggressiveness.** *Anticancer Res* 1993, **13(3)**:565-9
35. Friess H **Enhanced expression of transforming growth factor beta isoforms in pancreatic cancer correlates with decreased survival.** *Gastroenterology* 1993, **105(6)**:1846-56
36. Yokoyama MEM, Funatomi H, Friess H, Büchler MV, Johnson GR and Korc M **Amphiregulin is a potent mitogen in human pancreatic cancer cells: correlation with patient survival.** *Int J Oncol* 1995, **6**:625-631
37. Wagner M **Expression of a truncated EGF receptor is associated with inhibition of pancreatic cancer cell growth and enhanced sensitivity to cisplatin.** *Int J Cancer* 1996, **68(6)**:782-7
38. Matsuda K **Multiple mitogenic pathways in pancreatic cancer cells are blocked by a truncated epidermal growth factor receptor.** *Cancer Res* 2002, **62(19)**:5611-7
39. Wagner M **Suppression of fibroblast growth factor receptor signaling inhibits pancreatic cancer growth in vitro and in vivo.** *Gastroenterology* 1998, **114(4)**:798-807
40. Kornmann M, Arber N and Korc M **Inhibition of basal and mitogen-stimulated pancreatic cancer cell growth by cyclin D1 antisense is associated with loss of tumorigenicity and potentiation of cytotoxicity to cisplatin.** *J Clin Invest* 1998, **101(2)**:344-52
41. Overholser JP **Epidermal growth factor receptor blockade by antibody IMC-C225 inhibits growth of a human pancreatic carcinoma xenograft in nude mice.** *Cancer* 2000, **89(1)**:74-82
42. Bruns CJ **Blockade of the epidermal growth factor receptor signaling by a novel tyrosine kinase inhibitor leads to apoptosis of endothelial cells and therapy of human pancreatic carcinoma.** *Cancer Res* 2000, **60(11)**:2926-35
43. Ferrara N **Molecular and biological properties of vascular endothelial growth factor.** *J Mol Med* 1999, **77(7)**:527-43
44. Dvorak HF **Vascular permeability factor/vascular endothelial growth factor and the significance of microvascular hyperpermeability in angiogenesis.** *Curr Top Microbiol Immunol* 1999, **237**:97-132
45. Ortega N, Hutchings H and Plouet J **Signal relays in the VEGF system.** *Front Biosci* 1999, **4**:D141-52
46. Houck KA **The vascular endothelial growth factor family: identification of a fourth molecular species and characterization of alternative splicing of RNA.** *Mol Endocrinol* 1991, **5(12)**:1806-14
47. Poltorak Z **VEGF145, a secreted vascular endothelial growth factor isoform that binds to extracellular matrix.** *J Biol Chem* 1997, **272(11)**:7151-8
48. Park JE, Keller GA and Ferrara N **The vascular endothelial growth factor (VEGF) isoforms: differential deposition into the subepithelial extracellular matrix and bioactivity of extracellular matrix-bound VEGF.** *Mol Biol Cell* 1993, **4(12)**:1317-26
49. Olofsson B **Vascular endothelial growth factor B, a novel growth factor for endothelial cells.** *Proc Natl Acad Sci U S A* 1996, **93(6)**:2576-81
50. Grimmond S **Cloning and characterization of a novel human gene related to vascular endothelial growth factor.** *Genome Res* 1996, **6(2)**:124-31
51. Joukov V **A novel vascular endothelial growth factor, VEGF-C, is a ligand for the Flt4 (VEGFR-3) and KDR (VEGFR-2) receptor tyrosine kinases.** *Embo J* 1996, **15(7)**:1751
52. Yamada Y **Molecular cloning of a novel vascular endothelial growth factor, VEGF-D.** *Genomics* 1997, **42(3)**:483-8
53. Meyer M **A novel vascular endothelial growth factor encoded by Orf virus, VEGF-E, mediates angiogenesis via signalling through VEGFR-2 (KDR) but not VEGFR-1 (Flt-1) receptor tyrosine kinases.** *Embo J* 1999, **18(2)**:363-74
54. Maglione D **Isolation of a human placenta cDNA coding for a protein related to the vascular permeability factor.** *Proc Natl Acad Sci U S A* 1991, **88(20)**:9267-71
55. Ferrara N **Heterozygous embryonic lethality induced by targeted inactivation of the VEGF gene.** *Nature* 1996, **380(6573)**:439-42
56. Carmeliet P **Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele.** *Nature* 1996, **380(6573)**:435-9
57. Shibuya M **Structure and function of VEGF/VEGF-receptor system involved in angiogenesis.** *Cell Struct Funct* 2001, **26(1)**:25-35
58. Veikkola T **Regulation of angiogenesis via vascular endothelial growth factor receptors.** *Cancer Res* 2000, **60(2)**:203-12
59. Neufeld G **Vascular endothelial growth factor (VEGF) and its receptors.** *Faseb J* 1999, **13(1)**:9-22
60. Kukkk E **VEGF-C receptor binding and pattern of expression with VEGFR-3 suggests a role in lymphatic vascular development.** *Development* 1996, **122(12)**:3829-37
61. Ijijn K **VEGFR3 gene structure, regulatory region, and sequence polymorphisms.** *Faseb J* 2001, **15(6)**:1028-36
62. Soker S **Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor.** *Cell* 1998, **92(6)**:735-45
63. Kitsukawa T **Overexpression of a membrane protein, neuropilin, in chimeric mice causes anomalies in the cardiovascular system, nervous system and limbs.** *Development* 1995, **121(12)**:4309-18
64. Gluzman-Poltorak Z **Neuropilin-2 is a receptor for the vascular endothelial growth factor (VEGF) forms VEGF-145 and VEGF-165.** *J Biol Chem* 2000, **275(38)**:29922
65. Fong GH **Role of the Flt-1 receptor tyrosine kinase in regulating the assembly of vascular endothelium.** *Nature* 1995, **376(6535)**:66-70
66. Shalaby F **Failure of blood-island formation and vasculogenesis in Flk-1-deficient mice.** *Nature* 1995, **376(6535)**:62-6
67. Takashima S **Targeting of both mouse neuropilin-1 and neuropilin-2 genes severely impairs developmental yolk sac and embryonic angiogenesis.** *Proc Natl Acad Sci U S A* 2002, **99(6)**:3657-62
68. Hanahan D and Folkman J **Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis.** *Cell* 1996, **86(3)**:353-64
69. Fang J **HIF-1alpha-mediated up-regulation of vascular endothelial growth factor, independent of basic fibroblast growth factor, is important in the switch to the angiogenic phenotype during early tumorigenesis.** *Cancer Res* 2001, **61(15)**:5731-5
70. Giordano FJ and RS Johnson **Angiogenesis: the role of the microenvironment in flipping the switch.** *Curr Opin Genet Dev* 2001, **11(1)**:35-40
71. Udagawa T **Persistence of microscopic human cancers in mice: alterations in the angiogenic balance accompanies loss of tumor dormancy.** *Faseb J* 2002, **16(11)**:1361-70
72. Folkman J **What is the evidence that tumors are angiogenesis dependent?** *J Natl Cancer Inst* 1990, **82(1)**:4-6
73. Folkman J **Angiogenesis in cancer, vascular, rheumatoid and other disease.** *Nat Med* 1995, **1(1)**:27-31
74. Santin AD **Secretion of vascular endothelial growth factor in ovarian cancer.** *Eur J Gynaecol Oncol* 1999, **20(3)**:177-81
75. Plate KH **Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo.** *Nature* 1992, **359(6398)**:845-8
76. Goldman CK **Brain edema in meningiomas is associated with increased vascular endothelial growth factor expression.** *Neurosurgery* 1997, **40(6)**:1269-77
77. Okada F **Impact of oncogenes in tumor angiogenesis: mutant K-ras up-regulation of vascular endothelial growth factor/vascular permeability factor is necessary, but not sufficient for tumorigenicity of human colorectal carcinoma cells.** *Proc Natl Acad Sci U S A* 1998, **95(7)**:3609-14

78. Meadows KN, Bryant P and Pumiglia K **Vascular endothelial growth factor induction of the angiogenic phenotype requires Ras activation.** *J Biol Chem* 2001, **276(52)**:49289-98
79. Blancher C **Effects of ras and von Hippel-Lindau (VHL) gene mutations on hypoxia-inducible factor (HIF)-1alpha, HIF-2alpha, and vascular endothelial growth factor expression and their regulation by the phosphatidylinositol 3'-kinase/Akt signaling pathway.** *Cancer Res* 2001, **61(19)**:7349-55
80. Yu JL **Effect of p53 status on tumor response to antiangiogenic therapy.** *Science* 2002, **295(5559)**:1526-8
81. Shi Q **Constitutive Sp1 activity is essential for differential constitutive expression of vascular endothelial growth factor in human pancreatic adenocarcinoma.** *Cancer Res* 2001, **61(10)**:4143-54
82. Arora R **Angiogenesis as an independent prognostic indicator in node-negative breast cancer.** *Anal Quant Cytol Histol* 2002, **24(4)**:228-33
83. Sledge GW Jr **Vascular endothelial growth factor in breast cancer: biologic and therapeutic aspects.** *Semin Oncol* 2002, **29(3 Suppl 11)**:104-10
84. Millauer B **Glioblastoma growth inhibited in vivo by a dominant-negative Flk-1 mutant.** *Nature* 1994, **367(6463)**:576-9
85. Kong HL **Regional suppression of tumor growth by in vivo transfer of a cDNA encoding a secreted form of the extracellular domain of the flt-1 vascular endothelial growth factor receptor.** *Hum Gene Ther* 1998, **9(6)**:823-33
86. Goldman CK **Paracrine expression of a native soluble vascular endothelial growth factor receptor inhibits tumor growth, metastasis, and mortality rate.** *Proc Natl Acad Sci U S A* 1998, **95(15)**:8795-800
87. Kim KJ **Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo.** *Nature* 1993, **362(6423)**:841-4
88. Cheng SY **Suppression of glioblastoma angiogenicity and tumorigenicity by inhibition of endogenous expression of vascular endothelial growth factor.** *Proc Natl Acad Sci U S A* 1996, **93(16)**:8502-7
89. Saleh M, Stacker SA and Wilks AF **Inhibition of growth of C6 glioma cells in vivo by expression of antisense vascular endothelial growth factor sequence.** *Cancer Res* 1996, **56(2)**:393-401
90. Pavco PA **Antitumor and antimetastatic activity of ribozymes targeting the messenger RNA of vascular endothelial growth factor receptors.** *Clin Cancer Res* 2000, **6(5)**:2094-103
91. Fong TA **SU5416 is a potent and selective inhibitor of the vascular endothelial growth factor receptor (Flk-1/KDR) that inhibits tyrosine kinase catalysis, tumor vascularization, and growth of multiple tumor types.** *Cancer Res* 1999, **59(1)**:99-106
92. Witte L **Monoclonal antibodies targeting the VEGF receptor-2 (Flk1/KDR) as an anti-angiogenic therapeutic strategy.** *Cancer Metastasis Rev* 1998, **17(2)**:155-61
93. Seo Y **High expression of vascular endothelial growth factor is associated with liver metastasis and a poor prognosis for patients with ductal pancreatic adenocarcinoma.** *Cancer* 2000, **88(10)**:2239-45
94. Ikeda N **Prognostic significance of angiogenesis in human pancreatic cancer.** *Br J Cancer* 1999, **79**:9-10
95. Ellis LM **Vessel counts and vascular endothelial growth factor expression in pancreatic adenocarcinoma.** *Eur J Cancer* 1998, **34(3)**:337-40
96. Balaz P, Friess H and Büchler MW **Growth factors in pancreatic health and disease.** *Pancreatol* 2001, **1(4)**:343-55
97. Itakura J **Concomitant over-expression of vascular endothelial growth factor and its receptors in pancreatic cancer.** *Int J Cancer* 2000, **85(1)**:27-34
98. von Marschall Z **De novo expression of vascular endothelial growth factor in human pancreatic cancer: evidence for an autocrine mitogenic loop.** *Gastroenterology* 2000, **119(5)**:1358-72
99. Hotz HG **Angiogenesis inhibitor TNP-470 reduces human pancreatic cancer growth.** *J Gastrointest Surg* 2001, **5(2)**:131-8
100. Tokunaga T **Ribozyme mediated cleavage of cell-associated isoform of vascular endothelial growth factor inhibits liver metastasis of a pancreatic cancer cell line.** *Int J Oncol* 2002, **21(5)**:1027-32
101. Hotz HG **Specific targeting of tumor vasculature by diphtheria toxin-vascular endothelial growth factor fusion protein reduces angiogenesis and growth of pancreatic cancer.** *J Gastrointest Surg* 2002, **6(2)**:159-66
102. Hoshida T **Gene therapy for pancreatic cancer using an adenovirus vector encoding soluble flt-1 vascular endothelial growth factor receptor.** *Pancreas* 2002, **25(2)**:111-21
103. Ogawa T **Anti-tumor angiogenesis therapy using soluble receptors: enhanced inhibition of tumor growth when soluble fibroblast growth factor receptor-1 is used with soluble vascular endothelial growth factor receptor.** *Cancer Gene Ther* 2002, **9(8)**:633-40
104. Solorzano CC **Inhibition of growth and metastasis of human pancreatic cancer growing in nude mice by PTK 877/ZK22 an inhibitor of the vascular endothelial growth factor receptor tyrosine kinases.** *Cancer Biother Radiopharm* 2584, **16(5)**:359-70
105. Gupta VK **Vascular endothelial growth factor enhances endothelial cell survival and tumor radioresistance.** *Cancer J* 2002, **8(1)**:47-54
106. Harmey JH and Bouchier-Hayes D **Vascular endothelial growth factor (VEGF), a survival factor for tumour cells: implications for anti-angiogenic therapy.** *Bioessays* 2002, **24(3)**:280-3
107. Dias S **VEGF(165) promotes survival of leukemic cells by Hsp90-mediated induction of Bcl-2 expression and apoptosis inhibition.** *Blood* 2002, **99(7)**:2532-40
108. Gerber HP **VEGF regulates haematopoietic stem cell survival by an internal autocrine loop mechanism.** *Nature* 2002, **417(6892)**:954-8
109. Tang RF **Overexpression of lymphangiogenic growth factor VEGF-C in human pancreatic cancer.** *Pancreas* 2001, **22(3)**:285-92
110. Saimura M **Tumor suppression through angiogenesis inhibition by SUIT-2 pancreatic cancer cells genetically engineered to secrete NK4.** *Clin Cancer Res* 2002, **8(10)**:3243-9
111. Kleeff J **Detection and localization of Mip-3alpha/LARC/Exodus, a macrophage proinflammatory chemokine, and its CCR6 receptor in human pancreatic cancer.** *Int J Cancer* 1999, **81(4)**:650-7
112. Le X **Molecular regulation of constitutive expression of interleukin-8 in human pancreatic adenocarcinoma.** *J Interferon Cytokine Res* 2000, **20(11)**:935-46
113. Shi Q **Constitutive and inducible interleukin 8 expression by hypoxia and acidosis renders human pancreatic cancer cells more tumorigenic and metastatic.** *Clin Cancer Res* 1999, **5(11)**:3711-21
114. Massague J **TGF-beta signal transduction.** *Annu Rev Biochem* 1998, **67**:753-91
115. Rowland-Goldsmith MA **Soluble type II transforming growth factor-beta (TGF-beta) receptor inhibits TGF-beta signaling in COLO-357 pancreatic cancer cells in vitro and attenuates tumor formation.** *Clin Cancer Res* 2001, **7(9)**:2931-40
116. Rowland-Goldsmith MA, Matsuda K, Idezawa T, Ralli M, Ralli S and Korc M **Soluble type II transforming growth factor-beta receptor attenuates expression of metastasis-associated genes and suppresses pancreatic cancer cell metastasis.** *Mol Cancer Therapeutics* 2002, **1**:161-167
117. Takeuchi Y **Expression of plasminogen activators and their inhibitors in human pancreatic carcinoma: immunohistochemical study.** *Am J Gastroenterol* 1993, **88(11)**:1928-33
118. Kleeff J **Overexpression of Smad2 and colocalization with TGF-beta1 in human pancreatic cancer.** *Dig Dis Sci* 1999, **44(9)**:1793-802
119. Kleeff J and Korc M **Up-regulation of transforming growth factor (TGF)-beta receptors by TGF-beta1 in COLO-357 cells.** *J Biol Chem* 1998, **273(13)**:7495-500
120. Yang EY and Moses HL **Transforming growth factor beta 1-induced changes in cell migration, proliferation, and angiogenesis in the chicken chorioallantoic membrane.** *J Cell Biol* 1990, **111(2)**:731-41
121. Lambert V **Influence of plasminogen activator inhibitor type I on choroidal neovascularization.** *Faseb J* 2001, **15(6)**:1021-7
122. Andreasen PA, Egelund R and Petersen HH **The plasminogen activation system in tumor growth, invasion, and metastasis.** *Cell Mol Life Sci* 2000, **57(1)**:25-40
123. Munger JS **Latent transforming growth factor-beta: structural features and mechanisms of activation.** *Kidney Int* 1997, **51(5)**:1376-82

124. Kojima S, Nara K and Rifkin DB **Requirement for transglutaminase in the activation of latent transforming growth factor-beta in bovine endothelial cells.** *J Cell Biol* 1993, **121**(2):439-48
125. Cantero D **Enhanced expression of urokinase plasminogen activator and its receptor in pancreatic carcinoma.** *Br J Cancer* 1997, **75**(3):388-95
126. Ishiwata T **Altered expression of insulin-like growth factor II receptor in human pancreatic cancer.** *Pancreas* 1997, **15**(4):367-73
127. Elsasser HP **Characterization of a transglutaminase expressed in human pancreatic adenocarcinoma cells.** *Eur J Cell Biol* 1993, **61**(2):321-8
128. Haroon ZA **Tissue transglutaminase is expressed, active, and directly involved in rat dermal wound healing and angiogenesis.** *Faseb J* 1999, **13**(13):1787-95
129. Mishima K **A peptide derived from the non-receptor-binding region of urokinase plasminogen activator inhibits glioblastoma growth and angiogenesis in vivo in combination with cisplatin.** *Proc Natl Acad Sci U S A* 2000, **97**(15):8484-9
130. Schwarte-Waldhoff I **Smad4/DPC4-mediated tumor suppression through suppression of angiogenesis.** *Proc Natl Acad Sci U S A* 2000, **97**(17):9624-9
131. Liu D **EGFR is a transducer of the urokinase receptor initiated signal that is required for in vivo growth of a human carcinoma.** *Cancer Cell* 2002, **1**(5):445-57
132. Bancroft CC **Effects of pharmacologic antagonists of epidermal growth factor receptor, PI3K and MEK signal kinases on NF-kappaB and AP-1 activation and IL-8 and VEGF expression in human head and neck squamous cell carcinoma lines.** *Int J Cancer* 2002, **99**(4):538-48
133. Hirata A **ZD1839 (Iressa) induces antiangiogenic effects through inhibition of epidermal growth factor receptor tyrosine kinase.** *Cancer Res* 2002, **62**(9):2554-60
134. Tsuzuki Y **Pancreas microenvironment promotes VEGF expression and tumor growth: novel window models for pancreatic tumor angiogenesis and microcirculation.** *Lab Invest* 2001, **81**(10):1439-51
135. Yamanaka Y **Synthesis and expression of transforming growth factor beta-1, beta-2, and beta-3 in the endocrine and exocrine pancreas.** *Diabetes* 1993, **42**(5):746-56
136. Reinmuth N **Impact of insulin-like growth factor receptor-I function on angiogenesis, growth, and metastasis of colon cancer.** *Lab Invest* 2002, **82**(10):1377-89
137. Reinmuth N **Blockade of insulin-like growth factor I receptor function inhibits growth and angiogenesis of colon cancer.** *Clin Cancer Res* 2002, **8**(10):3259-69
138. Teraoka H **Enhanced VEGF production and decreased immunogenicity induced by TGF-beta 1 promote liver metastasis of pancreatic cancer.** *Br J Cancer* 2001, **85**(4):612-7
139. Ebert MP **Reduced PTEN expression in the pancreas overexpressing transforming growth factor-beta 1.** *Br J Cancer* 2002, **86**(2):257-62
140. Bergers G **Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis.** *Nat Cell Biol* 2000, **2**(10):737-44
141. Wen S **PTEN controls tumor-induced angiogenesis.** *Proc Natl Acad Sci U S A* 2001, **98**(8):4622-7
142. Liu CD **Vascular endothelial growth factor is increased in ascites from metastatic pancreatic cancer.** *J Surg Res* 2002, **102**(1):31-4
143. Niedergethmann M **High expression of vascular endothelial growth factor predicts early recurrence and poor prognosis after curative resection for ductal adenocarcinoma of the pancreas.** *Pancreas* 2002, **25**(2):122-9
144. Buchler P **VEGF-R11 Influences the Prognosis of Pancreatic Cancer.** *Ann Surg* 2002, **236**(6):738-49
145. Baker CH, Solorzano CC and Fidler IJ **Angiogenesis and cancer metastasis: antiangiogenic therapy of human pancreatic adenocarcinoma.** *Int J Clin Oncol* 2001, **6**(2):59-65

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

