

REVIEW

Open Access

Roles of Sema4D and Plexin-B1 in tumor progression

Ewe Seng Ch'ng^{1*}, Atsushi Kumanogoh²

Abstract

Sema4D, also known as CD100, is a protein belonging to class IV semaphorin. Its physiologic roles in the immune and nervous systems have been extensively explored. However, the roles of Sema4D have extended beyond these traditionally studied territories. Via interaction with its high affinity receptor Plexin-B1, Sema4D-Plexin-B1 involvement in tumor progression is strongly implied. Here, we critically review and delineate the Sema4D-Plexin-B1 interaction in many facets of tumor progression: tumor angiogenesis, regulation of tumor-associated macrophages and control of invasive growth. We correlate the in vitro and in vivo experimental data with the clinical study outcomes, and present a molecular mechanistic basis accounting for the intriguingly contradicting results from these recent studies.

1. Introduction

Semaphorins are a large family of secreted, transmembrane or glycosylphosphatidylinositol-linked proteins containing a phylogenetically conserved extracellular "sema" domain. They are classified into eight classes, of which classes 3 to 7 contain vertebrate semaphorins [1]. Their major receptors have been identified as plexins. In contrast to semaphorins, plexins are transmembrane proteins containing a phylogenetically conserved intracellular "sex and plexins" domain. They are subdivided into plexin A to D subfamilies [2].

Since the discovery of semaphorins as axon guidance cues, semaphorin-plexin interactions have been traditionally best described in the nervous system. Recently their interactions in other systems have also been explored, notably in the cardiovascular development, following revelation of their regulatory roles in cellular motility [3,4]. Intriguingly, semaphorins are also extensively involved in immune response regulation, qualifying some of them as "immune" semaphorins [5]. Recent studies have also focused on the pathological aspects, in particular on their links to several hallmarks of cancer such as tumor angiogenesis and invasiveness. Indeed, these processes recapitulate the many semaphorin-plexin roles in physiologic development. Several recent reviews

nicely summarized the accumulating evidence of semaphorin-plexin involvement in cancer progression [6-9]; however, the majority of the content referred to particularly secreted class 3 semaphorins. In this review, we focus on a transmembrane class 4 semaphorin, Sema4D and its receptor, plexin-B1. We also present a molecular mechanistic basis to reconcile the conflicting results of these molecules in tumor progression from the recent studies.

2. Sema4D has versatile roles in immune regulation and axonal guidance

Sema4D, also known as CD100, is a 150kD homodimeric transmembrane glycoprotein first identified in immune cells [10-12], and later classified as a member of class 4 semaphorin based on its distinctive structure [1].

Biological activities of Sema4D are well characterized in the immune system. Sema4D is highly expressed in resting T cells. Although its expression is low in resting B cells and antigen-presenting cells, Sema4D is up-regulated in these cells upon activation. As a ligand acting via its low affinity receptor CD72, Sema4D promotes the aggregation and survival of B cells, and enhances the activation of B cells in antibody production [11,13,14]. Sema4D is also involved in the activation and maturation of antigen-presenting cells [14,15]. Via its high affinity receptor Plexin-B1 [2], Sema4D inhibits the migration of monocytic and B-cell lineage cells [16,17]. Reciprocally, acting as a receptor, Sema4D regulates T

* Correspondence: chngeweseng@hotmail.com

¹Department of Pathology, School of Medical Sciences, Universiti Sains Malaysia, Malaysia

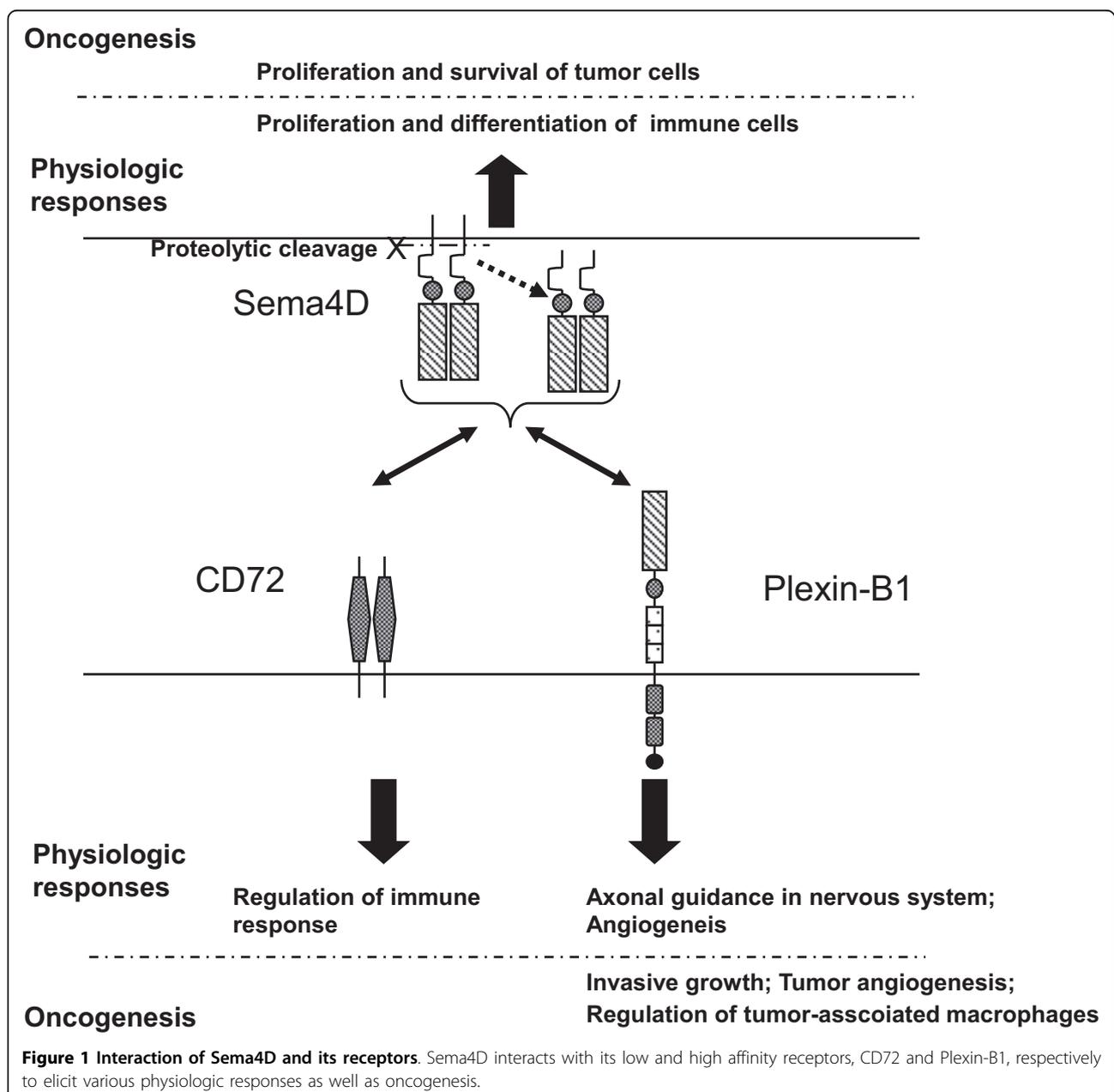
Full list of author information is available at the end of the article

cell activation [18,19], and is involved in the terminal stages of B cell differentiation [20].

Sema4D is also widely expressed in embryonic nervous tissue, especially in cortical plate and dorsal root ganglia [12], and postnatally in white matter, especially in myelinating oligodendrocytes [21]. As an axonal guidance cue in central nervous tissue, Sema4D functions in a seemingly contradicting fashion. For example, it induces growth cone collapse in primary hippocampal neurons [22,23], but stimulates axonal outgrowth of embryonic dorsal root ganglion neurons [24] and guides

the migratory process of gonadotropin hormone-releasing hormone-1 cells [25]. Contradictory to its interaction with CD72 in the immune system, Sema4D interacts with its high affinity receptor, Plexin-B1, in the nervous tissue.

However, cumulative data have suggested wider roles for Sema4D. In vitro and in vivo studies recently have given many insights into the involvement of Sema4D in tumor progression via interaction with Plexin-B1 (Figure 1). The mechanistic roles of Sema4D and Plexin-B1 in the tumor microenvironment such as tumor angiogenesis,



regulation of tumor-associated macrophages as well as the invasiveness of the tumor itself are highlighted in the following discussion.

3. *Sema4D* acts as an angiogenic factor through its high affinity receptor Plexin-B1

Evidence demonstrating the proangiogenic properties of *Sema4D* has emerged from several *in vitro* and *in vivo* studies. *Sema4D* is able to induce cell migration and promote tubulogenesis in endothelial cells, mimicking pivotal events in angiogenesis. Notably, this angiogenic response elicited by *Sema4D* is comparable to that by other well-known angiogenic molecules such as VEGF-A, HGF, and bFGF and is independent of any up-regulation of these molecules. *In vivo* studies employing chick chorioallantoic membrane (CAM) assay and Matigel plug assay confirmed this finding [26,27].

The signaling mechanism behind this *Sema4D*-induced angiogenic response has also been extensively studied. Plexin-B1, a high affinity receptor for *Sema4D*, is highly expressed in endothelial cells. *Sema4D* specifically binds and activates this endothelial receptor to elicit an angiogenic effect [26,27]. However, the signaling cascade beyond the *Sema4D*-Plexin-B1 interaction is complex and is still debated as described below.

Regarding the angiogenic process, two mechanisms have been proposed with conflicting results (Figure 2.). One mechanism proposes a signaling mechanism utilizing the C-terminal PDZ-binding motif of Plexin-B1, which binds two RhoGEFs, PDZ-RhoGEF and LARG, and subsequently activates RhoA. Through the engagement of integrins, this pathway also activates an intracellular tyrosine kinase cascade, notably one that involves Pyk2 [27-29]. The other proposed mechanism involves Met activation and tyrosine phosphorylation through Plexin-B1-Met interaction upon *Sema4D* binding [26]. Both of these mechanisms lead to cytoskeletal reorganization in endothelial cells and their migration.

Given its proangiogenic properties, the possible involvement of *Sema4D* in tumor angiogenesis was explored. *Sema4D* is indeed highly expressed in a wide range of human tumors such as prostate, colon, breast, oral, head and neck carcinomas as well as soft tissue sarcomas [30,31]. In one study employing head and neck squamous cell carcinoma cell lines with high expression of *Sema4D*, tumor growth and tumor angiogenesis *in vivo* were greatly impaired in the absence of *Sema4D* [31]. In addition, induction of angiogenesis by tumor cells was made possible by proteolytic cleavage of this membrane-bound *Sema4D* from the tumor cell surface, releasing its soluble form and thereby permitting *Sema4D* to act on local as well as distant tumor microenvironments [32].

Another study employing breast cancer cell lines in *Sema4D* knockout mice demonstrated that the

presence of *Sema4D* in the tumor microenvironment is important for tumor growth and metastasis, which is attributed to tumor vessel maturation induced by *Sema4D* [33]. Contradicting to the role of tumor-derived *Sema4D* in head and neck squamous cell carcinoma, this study however showed that tumor-associated macrophages but not tumor cells per se are the main cells that produce *Sema4D* in the tumor microenvironment, which enhances angiogenesis and contributes to tumor growth [33]. This discrepancy remains unsolved possibly due to differences between these tumor cell lines.

Similar to other *in vitro* studies, tumor angiogenesis induction by *Sema4D* is mediated through its interaction with Plexin-B1 [31] (Figure 2). A study using *Sema4D*-expressing melanoma transplants in Plexin-B1 knockout mice, however, showed no significant impairment in tumor angiogenesis. This result indicated that Plexin-B1 is sufficient but not essential for *Sema4D*-induced tumor angiogenesis; possibly the redundant lower affinity receptors such as Plexin-B2 could be engaged by *Sema4D* in tumor angiogenesis [34] (Figure 2).

4. *Sema4D* regulates monocytic lineage cells and tumor-associated macrophages in tumor microenvironment

Focusing on the effect of *Sema4D* on a monocytic lineage, an inhibitory effect on cell migration was demonstrated [16]. A recent study also showed that sequential engagement of CD72 and Plexin-B1 is crucial in this inhibitory effect during the maturation of monocytes into immature dendritic cells. Modulation of cytokine production towards either an anti-inflammatory or a pro-inflammatory cytokine profile seems to be dependent on *Sema4D* concentration [14,17]. Upon activation, *Sema4D* expression in dendritic cells is up-regulated. Augmentation of response in an autocrine manner by the proteolytic release of *Sema4D* acting on receptors in the same cells could ensue [15]. In tumor-associated macrophages, although the lack of *Sema4D* itself did not alter the differentiation and activation capabilities as well as the cytokine production profile, a significantly lower number of tumor-associated macrophages was observed in the tumor microenvironment [33]. However, the precise mechanism behind these findings awaits elucidation.

Considering these results, the inhibition of cell migration by *Sema4D* could be alternately interpreted as a means of prolonging cell-cell contact in a physiologic immune response. Exploitation of this mechanism by tumor cells would lead to the retention of tumor-associated macrophages and the subsequent induction of angiogenesis could therefore be triggered.

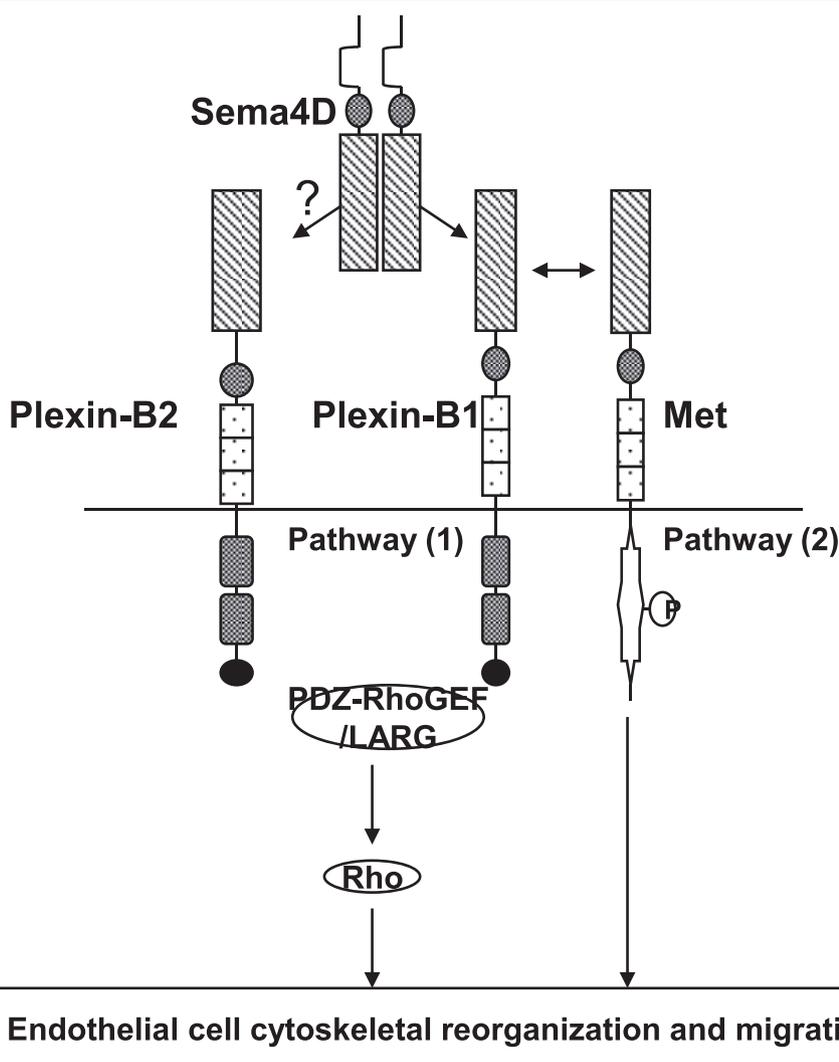


Figure 2 Sema4D-Plexin-B1 signaling pathways in angiogenesis. Sema4D engages Plexin-B1 to induce angiogenesis and tumor angiogenesis via two independent downstream pathways. Plexin-B2 might also play a redundant role in this aspect.

5. Sema4D-Plexin-B1 interaction controls invasive growth

Invasive growth is a complex morphogenetic program that includes several steps such as cell-cell dissociation, motility, colonization of new sites, proliferation and survival. Invasive growth under physiologic conditions occurs during embryogenesis and wound healing. However, when this program is hijacked, cancer progression and metastasis, the pathologic counterpart, ensue [35]. An invasive growth phenotype in epithelial cells could be elicited by Sema4D through Plexin-B1 coupled with the Met signaling pathway [36]. Another study along the same line showed that Ron, a scatter factor receptor similar to Met, also couples with Plexin-B1 in transducing Sema4D signals (Figure 3). Moreover, over-expression of Plexin-B1 is observed in breast, colon, liver,

pancreatic and gastric carcinoma cell lines. Signaling through Plexin-B1 coupled with Met is essential for invasive response in these cell lines [26,37,38].

However, signaling by Sema4D through Plexin-B1 coupling with Met in invasive response is partially disputed by a recent study. This study demonstrated that the antimigratory or promigratory effects of Sema4D in breast cancer cell lines depend on the stoichiometry of the expression of Plexin-B1, Met and ErbB-2 [39]. ErbB-2 is a well-known poor prognostic factor in breast cancer. Coupling of Plexin-B1 with Met inhibits migration, while coupling with ErbB-2 promotes it. By competing for Plexin-B1, these two molecules counter each other's effects in the same tumor cell, determining the net antimigratory or promigratory effect of Sema4D. These molecular complexes further control a downstream

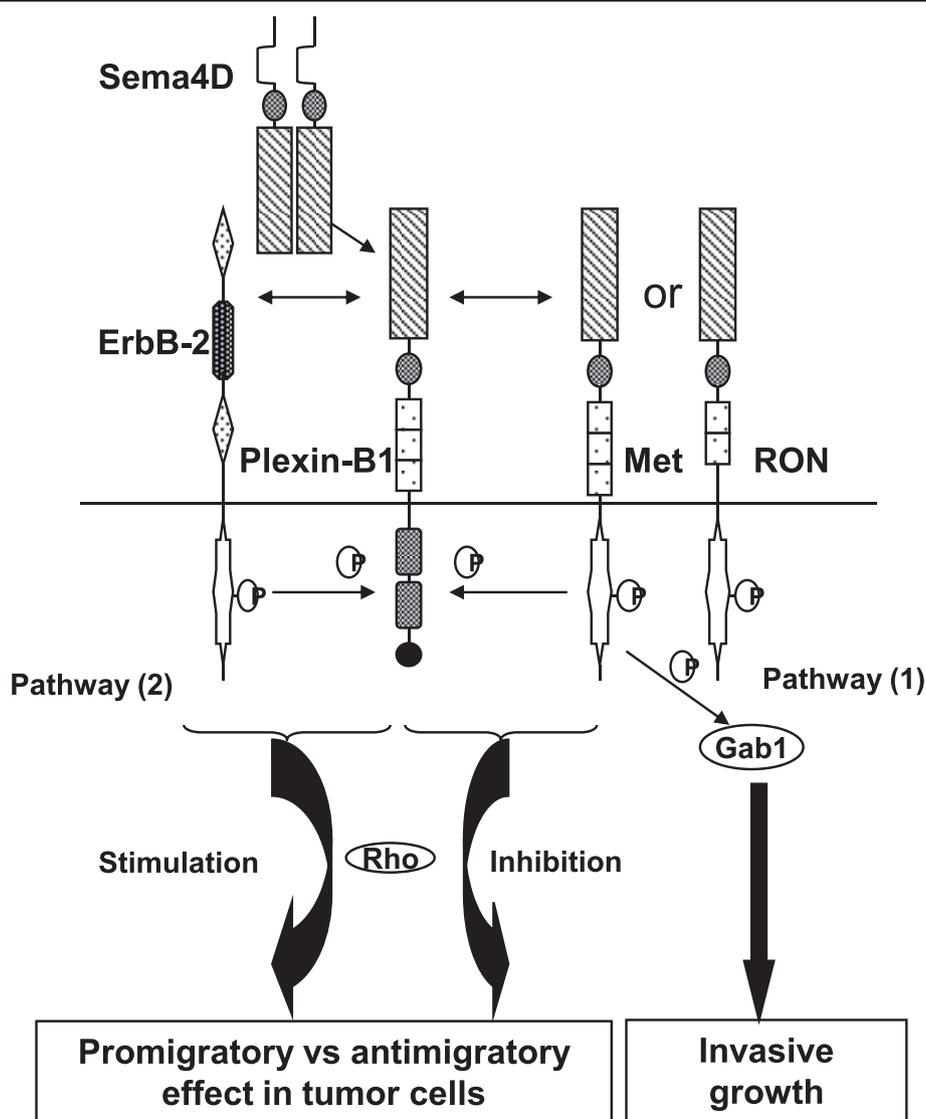


Figure 3 Control of cellular motility by Sema 4D-Plexin-B1 interaction via the cooperative mechanism. Coupled with the Met, Ron or ErbB-2 tyrosine kinase receptor at the cellular membrane surface, Sema4D interacts with Plexin-B1 to control cellular motility and invasive growth via Gab1 or Rho-dependant pathways.

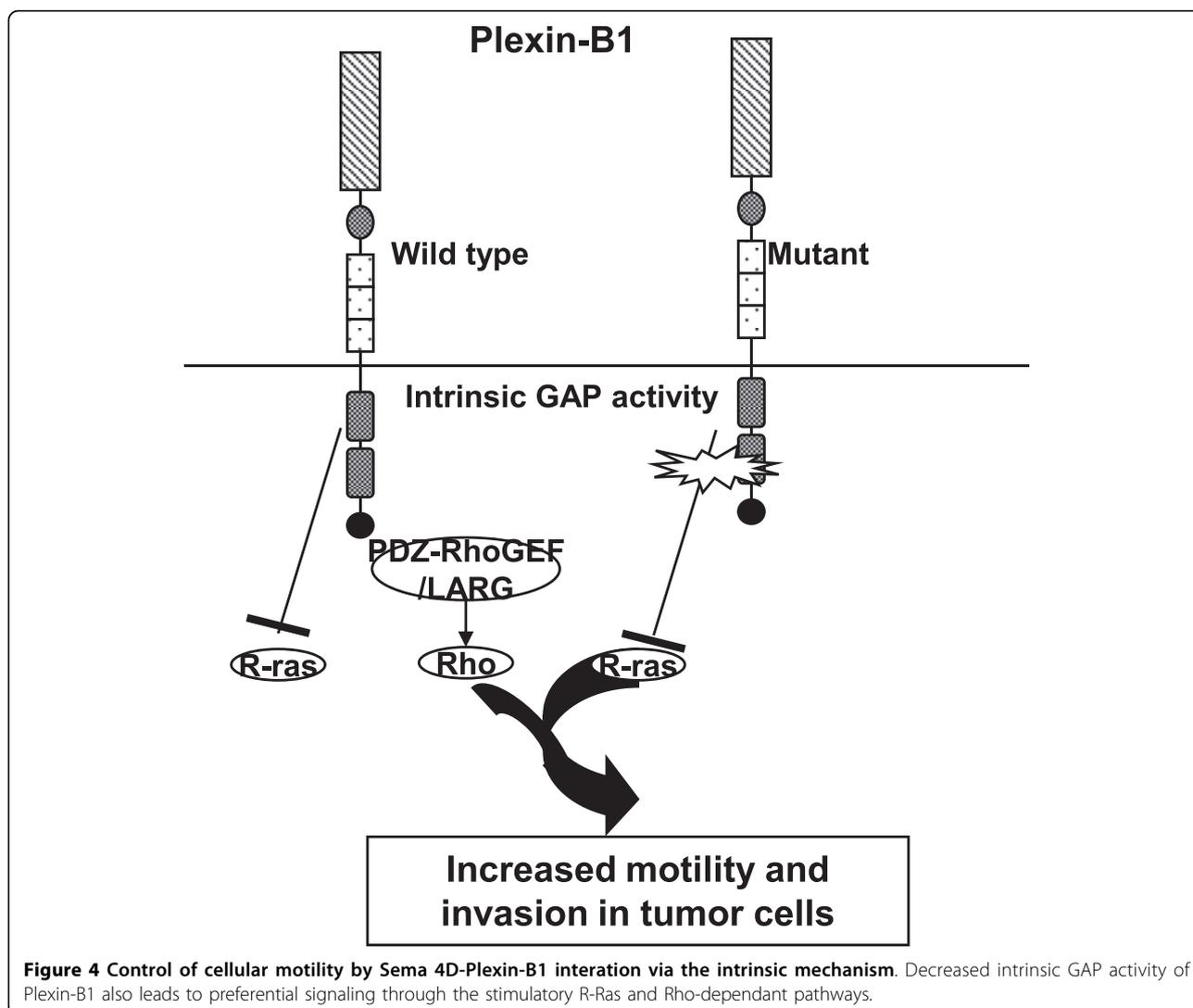
Rho-dependent pathway that is similar to the mechanism proposed to promote endothelial cell angiogenesis [39,40] (Figure 3).

Although not shown in most studies involving cancer cell lines, another line of investigation of Sema4D and Plexin-B1 interactions centered on the collapse of neurite outgrowth. Recently, Plexin-B1 has been discovered to have intrinsic GTPase-activating protein (GAP) activity towards R-Ras [23]. Inactivation of R-Ras reduces cell adhesion, cell migration, and growth cone outgrowth [41]. In contrast to the activating pathways discussed above, this inhibitory pathway is believed to inhibit cancer cell migration. Mutations that abolish the

R-Ras GAP activity of Plexin-B1 in cell lines promote cell adhesion, motility and invasion [42] (Figure 4).

6. High expression of Sema4D is a poor prognostic factor in soft tissue sarcomas

Cumulative evidence regarding the roles of Sema4D-Plexin-B1 interaction in tumor progression came from in vitro and in vivo studies. A recent clinical study on soft tissue sarcomas revealed that higher Sema4D expression was correlated with higher mitotic counts, cellularity, a higher necrosis ratio and Ki-67 index [30], suggesting a proliferative advantage in tumors with higher Sema4D expression. Accordingly, higher Sema4D



expression is associated with poorer overall survival and disease-free survival [30] (Table 1). Similarly, via activation by Plexin-B1, Sema4D-expressing CD38+ B-cell chronic lymphocytic leukemic cells have better viability and a higher proliferative rate [43].

7. Clinical studies offer insights into the role of Plexin-B1 in human oncogenesis

Recently clinical studies have also provided further insights into the significance of Plexin-B1 in human cancers (Table 1.). In human breast cancers, gene expression microarray analysis showed that low Plexin-B1 expression was associated with high histologic grading, ErbB-2 over-expression and a high proliferative index in estrogen receptor-positive breast cancers. Low Plexin-B1 expression also predicts poor outcome in terms of disease-free survival in estrogen receptor-positive breast cancers [44,45]. Similarly, Plexin-B1

expression is also down-regulated in renal cell carcinomas where 169 out of 209 carcinomas showed negativity in a recent report. Reintroducing Plexin-B1 into the renal adenocarcinoma cell line reduced the proliferative rate [46]. These results imply a tumor suppressor role of Plexin-B1 in oncogenesis.

In contrast, a recent study on human breast and ovarian cancers using immunohistochemistry showed that co-expression of Plexin-B1 and Met was significantly associated with a higher tumor grade and lymph node metastasis. Met was activated in the presence of Plexin-B1 expression in a majority of these cancers [47]. In human prostate cancers, Plexin-B1 and Sema4D are strongly associated and highly expressed compared to the non-neoplastic tissue (77% versus 6% and 58% versus 3.5% respectively). Mutations in Plexin-B1 are frequently detected in primary and metastatic prostate cancers [42]. Among the mutations, the more frequent

Table 1 Important findings regarding expression of *Sema4D* and *Plexin-B1* in various human cancers

Cancer Type	Findings	Prognosis
Soft tissue sarcomas [30]	Higher <i>Sema4D</i> expression was associated with higher mitotic counts, cellularity, necrotic ratio and Ki-67 index	Poorer overall survival and disease-free survival with higher <i>Sema4D</i> expression
B-cell chronic lymphocytic leukemia [43]	<i>Sema4D</i> -expressing CD38+ B-CLL cells showed increased proliferation and survival.	Not available
Breast carcinoma [44,45]	Lower <i>Plexin-B1</i> was associated with a higher histologic grade, proliferative index and <i>ErbB-2</i> expression in estrogen receptor-positive cases	Poorer disease-free survival with lower <i>Plexin-B1</i> expression in estrogen receptor- positive cases
Renal cell carcinoma [46]	81% of renal cell carcinoma lost the <i>plexin-B1</i> expression	Not available
Breast and Ovarian Carcinomas [47]	Co-expression of <i>Plexin-B1</i> and <i>Met</i> resulted in <i>Met</i> activation and was associated with a higher tumor grade and lymph node metastasis	Not available
Prostate adenocarcinoma [42]	Expression of <i>Plexin-B1</i> and <i>Sema4D</i> was significantly correlated and both were highly expressed in cancer cases (77% and 58% respectively)	Not available

amino acid changes are mapped to the Rho GTPase binding domain of *Plexin-B1*. Structural analysis revealed a reduced Rho GTPase binding affinity in some of these mutations [48]. In vitro studies showed that these mutations impaired the R-RasGAP activity of *Plexin-B1*, leading to increased cell adhesion, motility and invasion [42]. These results instead suggest an oncogenic nature of *Plexin-B1*. Interestingly, a recent study on melanoma also revealed contradicting effects of *Plexin-B1* expression on cell migration: enhancement in primary tumor cells but inhibition in metastatic tumor cells [49].

8. Intrinsic and cooperative mechanisms determine the overall signaling of *Plexin-B1* in tumor progression

Recognizing the fact that both the stimulatory and inhibitory domains coexist in the *Plexin-B1* cytoplasmic domain, we speculate that “intra-molecular” preferential *Plexin-B1* signaling pathways in cancers could constitute a sound explanation to reconcile these conflicting clinical results of *Plexin-B1* in tumor progression. Inhibitory signals of *Plexin-B1* would enable it to function as a tumor suppressor gene as implied from the breast and renal cancer studies [44-46]. In contrast, impairment in the *Plexin-B1* inhibitory pathway due to mutation, together with over-expression of *Plexin-B1* would allow its stimulatory signals to prevail and act as an oncogene such as in prostate cancer [42].

In addition, fine-tuning the inhibitory or stimulatory downstream signaling of *Plexin-B1* is possible through the association with competing receptor tyrosine kinases such as *Met* and *ErbB-2*, which in turn would deliver additional signals. Although a recent study on human breast and ovarian cancers implies that *Plexin-B1* functions as an oncogene when coupled with *Met* activation [47], involvement of *ErbB-2* in this association has yet

to be addressed in clinical studies. The available in vitro data, nonetheless, support the *ErbB-2* instead of *Met* coupling in determining the invasiveness of tumor cell lines [39,40].

Conclusion and perspective

Although the interaction of *Sema4D* and *Plexin-B1* has profound physiologic effects in the immune and nervous systems, emerging data suggests their involvement in tumor progression and especially in tumor angiogenesis where *Sema4D* acts as a proangiogenic factor via *Plexin-B1*. Over-expression of *Sema4D* in tumor confers on the tumor cells a selective proliferative advantage. *Sema4D* might also influence via *Plexin-B1* several functional aspects of tumor-associated macrophages such as their migratory capability and capability to induce tumor angiogenesis. In contrast to responses elicited in non-tumorous components such as endothelial cells and macrophages, signaling through *Plexin-B1* in various cancers gives rise to seemingly contradicting responses, as highlighted in this review. Current results suggest that either the stimulatory or inhibitory signaling could be induced via the respective stimulatory or inhibitory domain in the *Plexin-B1* molecule itself. This could also be achieved via association with the competing receptor tyrosine kinases such as *Met* and *ErbB-2* (Figure 3 and 4). This double-checkpoint mechanism based on intrinsic and cooperative signaling signifies the complexity and importance of *Plexin-B1* signaling in tumor progression. Further studies will certainly solidify the basis for these unique mechanisms.

List of abbreviations

VEGF: vascular endothelial growth factor; HGF: hepatocyte growth factor; bFGF: basic fibroblast growth factor; PDZ: PSD-95/Dlg/ZO-1; GEF: guanine nucleotide exchange factor; LARG: leukemia-associated Rho GEF; Pyk2:

Proline-rich tyrosine kinase 2; MET: mesenchymal-epithelial transition factor; Ron: Recepteur d'origine Nantais; GAP: GTPase-activating protein.

Acknowledgements

We thank Dr. Aik Hong Teh for his help in revision of the manuscript.

Author details

¹Department of Pathology, School of Medical Sciences, Universiti Sains Malaysia, Malaysia. ²Department of Immunopathology, Immunology Frontier Research Center, Research Institute for Microbial Diseases, Osaka University, Japan.

Authors' contributions

ESC drafted the manuscript. All authors analyzed the published data, read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 6 March 2010 Accepted: 21 September 2010

Published: 21 September 2010

References

- Goodman CS, Kolodkin AL, Luo Y, Püschel AW, Raper JA: **Unified Nomenclature for the Semaphorins/Collapsins.** *Cell* 1999, **97**:551-552.
- Tamagnone L, Artigiani S, Chen H, He Z, Ming G-I, Song H-j, Chedotal A, Winberg ML, Goodman CS, Poo M-m, et al: **Plexins Are a Large Family of Receptors for Transmembrane, Secreted, and GPI-Anchored Semaphorins in Vertebrates.** *Cell* 1999, **99**:71-80.
- Pasterkamp RJ, Giger RJ: **Semaphorin function in neural plasticity and disease.** *Curr Opin Neurobiol* 2009, **19**:263-274.
- Tran TS, Kolodkin AL, Bharadwaj R: **Semaphorin Regulation of Cellular Morphology.** *Annu Rev Cell Dev Biol* 2007, **23**:263-292.
- Mizui M, Kumanogoh A, Kikutani H: **Immune Semaphorins: Novel Features of Neural Guidance Molecules.** *J Clin Immunol* 2009, **29**:1-11.
- Neufeld G, Kessler O: **The semaphorins: versatile regulators of tumour progression and tumour angiogenesis.** *Nat Rev Cancer* 2008, **8**:632-645.
- Capparuccia L, Tamagnone L: **Semaphorin signaling in cancer cells and in cells of the tumor microenvironment - two sides of a coin.** *J Cell Sci* 2009, **122**:1723-1736.
- Flannery E, Duman-Scheel M: **Semaphorins at the interface of development and cancer.** *Curr Drug Targets* 2009, **10**:611-619.
- Potiron VA, Roche J, Drabkin HA: **Semaphorins and their receptors in lung cancer.** *Cancer Lett* 2009, **273**:1-14.
- Bougeret C, Mansur IG, Dastot H, Schmid M, Mahouy G, Bensussan A, Bomsell L: **Increased surface expression of a newly identified 150-kDa dimer early after human T lymphocyte activation.** *J Immunol* 1992, **148**:318-323.
- Hall KT, Bomsell L, Schultze JL, Boussiotis VA, Dorfman DM, Cardoso AA, Bensussan A, Nadler LM, Freeman GJ: **Human CD100, a novel leukocyte semaphorin that promotes B-cell aggregation and differentiation.** *Proc Natl Acad Sci USA* 1996, **93**:11780-11785.
- Furuyama T, Inagaki S, Kosugi A, Noda S, Saitoh S-i, Ogata M, Iwahashi Y, Miyazaki N, Hamaoka T, Tohyama M: **Identification of a Novel Transmembrane Semaphorin Expressed on Lymphocytes.** *J Biol Chem* 1996, **271**:33376-33381.
- Kumanogoh A, Watanabe C, Lee I, Wang X, Shi W, Araki H, Hirata H, Iwahori K, Uchida J, Yasui T, et al: **Identification of CD72 as a Lymphocyte Receptor for the Class IV Semaphorin CD100: A Novel Mechanism for Regulating B Cell Signaling.** *Immunity* 2000, **13**:621-631.
- Ishida I, Kumanogoh A, Suzuki K, Akahani S, Noda K, Kikutani H: **Involvement of CD100, a lymphocyte semaphorin, in the activation of the human immune system via CD72: implications for the regulation of immune and inflammatory responses.** *Int Immunol* 2003, **15**:1027-1034.
- Kumanogoh A, Suzuki K, Ch'ng E, Watanabe C, Marukawa S, Takegahara N, Ishida I, Sato T, Habu S, Yoshida K, et al: **Requirement for the lymphocyte semaphorin, CD100, in the induction of antigen-specific T cells and the maturation of dendritic cells.** *J Immunol* 2002, **169**:1175-1181.
- Delaire S, Billard C, Tordjman R, Chedotal A, Elhabazi A, Bensussan A, Bomsell L: **Biological Activity of Soluble CD100. II. Soluble CD100, Similarly to H-SemaIII, Inhibits Immune Cell Migration.** *J Immunol* 2001, **166**:4348-4354.
- Chabbert-de Ponnat I, Marie-Cardine A, Pasterkamp RJ, Schiavon V, Tamagnone L, Thomasset N, Bensussan A, Bomsell L: **Soluble CD100 functions on human monocytes and immature dendritic cells require plexin C1 and plexin B1, respectively.** *Int Immunol* 2005, **17**:439-447.
- Herold C, Bismuth G, Bensussan A, Bomsell L: **Activation signals are delivered through two distinct epitopes of CD100, a unique 150 kDa human lymphocyte surface structure previously defined by BB18 mAb.** *Int Immunol* 1995, **7**:1-8.
- Herold C, Elhabazi A, Bismuth G, Bensussan A, Bomsell L: **CD100 is associated with CD45 at the surface of human T lymphocytes. Role in T cell homotypic adhesion.** *J Immunol* 1996, **157**:5262-5268.
- Billard C, Delaire S, Raffoux E, Bensussan A, Bomsell L: **Switch in the protein tyrosine phosphatase associated with human CD100 semaphorin at terminal B-cell differentiation stage.** *Blood* 2000, **95**:965-972.
- Moreau-Fauvarque C, Kumanogoh A, Camand E, Jaillard C, Barbin G, Boquet I, Love C, Jones EY, Kikutani H, Lubetzki C, et al: **The Transmembrane Semaphorin Sema4D/CD100, an Inhibitor of Axonal Growth, Is Expressed on Oligodendrocytes and Upregulated after CNS Lesion.** *J Neurosci* 2003, **23**:9229-9239.
- Swiercz JM, Kuner R, Behrens J, Offermanns S: **Plexin-B1 Directly Interacts with PDZ-RhoGEF/LARG to Regulate RhoA and Growth Cone Morphology.** *Neuron* 2002, **35**:51-63.
- Oinuma I, Ishikawa Y, Katoh H, Negishi M: **The Semaphorin 4D receptor Plexin-B1 is a GTPase activating protein for R-Ras.** *Science* 2004, **305**:862-865.
- Kenta M, Tatsuo F, Mizue T, Shiho F, Hitomi K, Shinobu I: **Sema4D stimulates axonal outgrowth of embryonic DRG sensory neurones.** *Genes Cells* 2004, **9**:821-829.
- Giacobini P, Messina A, Morello F, Ferraris N, Corso S, Penachioni J, Giordano S, Tamagnone L, Fasolo A: **Semaphorin 4D regulates gonadotropin hormone-releasing hormone-1 neuronal migration through PlexinB1-Met complex.** *J Cell Biol* 2008, **183**:555-566.
- Conrotto P, Valdembrì D, Corso S, Serini G, Tamagnone L, Comoglio PM, Bussolino F, Giordano S: **Sema4D induces angiogenesis through Met recruitment by Plexin B1.** *Blood* 2005, **105**:4321-4329.
- Basile JR, Barac A, Zhu T, Guan K-L, Gutkind JS: **Class IV Semaphorins Promote Angiogenesis by Stimulating Rho-Initiated Pathways through Plexin-B.** *Cancer Res* 2004, **64**:5212-5224.
- Basile JR, Gavard J, Gutkind JS: **Plexin-B1 Utilizes RhoA and Rho Kinase to Promote the Integrin-dependent Activation of Akt and ERK and Endothelial Cell Motility.** *J Biol Chem* 2007, **282**:34888-34895.
- Basile JR, Afkhami T, Gutkind JS: **Semaphorin 4D/Plexin-B1 Induces Endothelial Cell Migration through the Activation of PYK2, Src, and the Phosphatidylinositol 3-Kinase-Akt Pathway.** *Mol Cell Biol* 2005, **25**:6889-6898.
- Ch'ng E, Tomita Y, Zhang B, He J, Hoshida Y, Qiu Y, Morii E, Nakamichi I, Hamada K, Ueda T, Aozasa K: **Prognostic significance of CD100 expression in soft tissue sarcoma.** *Cancer* 2007, **110**:164-172.
- Basile JR, Castilho RM, Williams VP, Gutkind JS: **Semaphorin 4D provides a link between axon guidance processes and tumor-induced angiogenesis.** *Proc Natl Acad Sci USA* 2006, **103**:9017-9022.
- Basile JR, Holmbeck K, Bugge TH, Gutkind JS: **MT1-MMP Controls Tumor-induced Angiogenesis through the Release of Semaphorin 4D.** *J Biol Chem* 2007, **282**:6899-6905.
- Sierra JR, Corso S, Caione L, Cepero V, Conrotto P, Cignetti A, Piacibello W, Kumanogoh A, Kikutani H, Comoglio PM, et al: **Tumor angiogenesis and progression are enhanced by Sema4D produced by tumor-associated macrophages.** *J Exp Med* 2008, **205**:1673-1685.
- Fazzari P, Penachioni J, Gianola S, Rossi F, Eickholt B, Maina F, Alexopoulou L, Sottile A, Comoglio P, Flavell R, Tamagnone L: **Plexin-B1 plays a redundant role during mouse development and in tumour angiogenesis.** *BMC Dev Biol* 2007, **7**:55.
- Trusolino L, Comoglio PM: **Scatter-factor and semaphorin receptors: cell signalling for invasive growth.** *Nat Rev Cancer* 2002, **2**:289-300.
- Giordano S, Corso S, Conrotto P, Artigiani S, Gilestro G, Barberis D, Tamagnone L, Comoglio PM: **The Semaphorin 4D receptor controls invasive growth by coupling with Met.** *Nat Cell Biol* 2002, **4**:720-724.

37. Conrotto P, Corso S, Gamberini S, Comoglio PM, Giordano S: **Interplay between scatter factor receptors and B plexins controls invasive growth.** *Oncogene* 2004, **23**:5131-5137.
38. Artigiani S, Barberis D, Fazzari P, Longati P, Angelini P, van de Loo J-W, Comoglio PM, Tamagnone L: **Functional Regulation of Semaphorin Receptors by Proprotein Convertases.** *J Biol Chem* 2003, **278**:10094-10101.
39. Swiercz JM, Worzfeld T, Offermanns S: **ErbB-2 and Met Reciprocally Regulate Cellular Signaling via Plexin-B1.** *J Biol Chem* 2008, **283**:1893-1901.
40. Swiercz JM, Kuner R, Offermanns S: **Plexin-B1/RhoGEF-mediated RhoA activation involves the receptor tyrosine kinase ErbB-2.** *J Cell Biol* 2004, **165**:869-880.
41. Oinuma I, Katoh H, Negishi M: **Semaphorin 4D/Plexin-B1-mediated R-Ras GAP activity inhibits cell migration by regulating β_1 integrin activity.** *J Cell Biol* 2006, **173**:601-613.
42. Wong OG, Nitkunan T, Oinuma I, Zhou C, Blanc V, Brown RS, Bott SR, Nariculam J, Box G, Munson P, et al: **Plexin-B1 mutations in prostate cancer.** *Proc Natl Acad Sci USA* 2007, **104**:19040-19045.
43. Deaglio S, Vaisitti T, Bergui L, Bonello L, Horenstein AL, Tamagnone L, Boumsell L, Malavasi F: **CD38 and CD100 lead a network of surface receptors relaying positive signals for B-CLL growth and survival.** *Blood* 2005, **105**:3042-3050.
44. Rody A, Karn T, Ruckhaberle E, Hanker L, Metzler D, Muller V, Solbach C, Ahr A, Gaetje R, Holtrich U, Kaufmann M: **Loss of Plexin B1 is highly prognostic in low proliferating ER positive breast cancers—results of a large scale microarray analysis.** *Eur J Cancer* 2009, **45**:405-413.
45. Rody A, Holtrich U, Gaetje R, Gehrmann M, Engels K, von Minckwitz G, Loibl S, Diallo-Danebrock R, Ruckhaberle E, Metzler D, et al: **Poor outcome in estrogen receptor-positive breast cancers predicted by loss of plexin B1.** *Clin Cancer Res* 2007, **13**:1115-1122.
46. Gomez Roman JJ, Garay GO, Saenz P, Escuredo K, Sanz Ibayondo C, Gutkind S, Junquera C, Simon L, Martinez A, Fernandez Luna JL, Val-Bernal JF: **Plexin B1 is downregulated in renal cell carcinomas and modulates cell growth.** *Transl Res* 2008, **151**:134-140.
47. Valente G, Nicotra G, Arrondini M, Castino R, Capparuccia L, Prat M, Kerim S, Tamagnone L, Isidoro C: **Co-expression of plexin-B1 and Met in human breast and ovary tumours enhances the risk of progression.** *Cell Oncol* 2009, **31**:423-436.
48. Tong Y, Hota PK, Hamaneh MB, Buck M: **Insights into Oncogenic Mutations of Plexin-B1 Based on the Solution Structure of the Rho GTPase Binding Domain.** *Structure* 2008, **16**:246-258.
49. Argast GM, Croy CH, Coutts KL, Zhang Z, Litman E, Chan DC, Ahn NG: **Plexin B1 is repressed by oncogenic B-Raf signaling and functions as a tumor suppressor in melanoma cells.** *Oncogene* 2009, **28**:2697-2709.

doi:10.1186/1476-4598-9-251

Cite this article as: Ch'ng and Kumanogoh: Roles of Sema4D and Plexin-B1 in tumor progression. *Molecular Cancer* 2010 **9**:251.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

